UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL FACULDADE DE ODONTOLOGIA

Programa de Pós-Graduação em Odontologia Doutorado em Odontologia Área de Concentração Clínica Odontológica – Materiais Dentários

CIMENTO EXPERIMENTAL BIOATIVO À BASE DE GLICEROL SALICILATO CONTENDO α-FOSFATO TRICÁLCICO PARA USO ENDODÔNTICO

Fernando Freitas Portella

Tese apresentada como requisito obrigatório para obtenção do título de **Doutor em Odontologia** na área de concentração em Clínica Odontológica.

Susana Maria Werner Samuel
Orientadora

Porto Alegre, março de 2015

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL FACULDADE DE ODONTOLOGIA PROGRAMA DE PÓS-GRADUAÇÃO EM ODONTOLOGIA

Porto Alegre, 24 de março de 2015

Ao PPGODO-ODO

Vimos por meio deste informar que as sugestões de alteração propostas pela Banca examinadora da Tese de Doutorado do aluno **Fernando Freitas Portella**, composta pelos professores Mario Tanomaru Filho (UNESP Araraquara), Antonio Shigueaki Takimi (UFPel) e Fabiana Soares Grecca (UFRGS), defendida às 8h30min do dia 23 de março de 2015, foram discutidas entre o orientador e o doutorando, e a Tese foi editada considerando os aspectos debatidos.

Sem mais para o momento.

Atenciosamente.

Prof. Dra. Susana Maria Werner Samuel

Ilmo.Prof. Dr. Cassiano Cassiano Kuchenbecker Rosing Coordenador do Programa de Pós-Graduação em Odontologia Nesta Faculdade

"A vontade de se preparar tem que ser maior do que a vontade de vencer. Vencer será consequência da boa preparação."

Bernardo Rezende

DEDICATÓRIA

Dedico este trabalho ao meu avô **Antônio Cândido de Freitas**, o impulsionador do estudo na minha família. De pouco estudo, mas de sabedoria infindável.

AGRADECIMENTOS

Aos meus pais **Moacir** e **Sandra** e ao meu irmão **Marcele** por me proporcionarem a melhor condição para estudar, pelo incentivo e acima de tudo por me mostrarem os melhores valores que uma pessoa pode ter.

À **Daniela Izolan** por me fazer entender melhor a vida, me mostrar a subjetividade indispensável à felicidade, me ensinar o que é o amor.

Aos meus amigos **Fernando Rios**, **Luciana Daudt** e **Ricardo Costa** pela memorável convivência nesses 10 anos de UFRGS.

À professora **Susana Maria Werner Samuel**, minha orientadora há 9 anos. Obrigado pelas oportunidades e confiança desde os tempos de PET, pelos ensinamentos científicos e principalmente pelo exemplo de pessoa que és para mim.

Aos professores **Fabrício Mezzomo Collares** e **Vicente Castelo Branco Leitune**, pela convivência, parceria, ensinamentos e cobrança, que certamente me fizeram um profissional melhor.

Aos amigos do Laboratório de Materiais Dentários (LAMAD), representados pela professora Carmen Fortes, pela convivência e colaboração durante todos os meus anos de LAMAD. Um agradecimento especial às alunas de iniciação científica Paula Dapper e Tamy Kowalski pela contribuição no desenvolvimento dos experimentos dessa tese.

Ao Laboratório de Biomateriais e Cerâmicas Avançadas (LabioMat), na pessoa do professor Luis Alberto dos Santos; ao Laboratório de Síntese e Polímeros (SinPol), na pessoa do professor Cesar Liberato Petzhold; e ao Laboratório de Células-tronco e Engenharia de Tecidos da Universidade Luterana do Brasil, na pessoa da professora Melissa Camassola, pela parceria na realização desse estudo.

Ao Programa de Pós-Graduação em Odontologia pela oportunidade de realizar o Doutorado e pela convivência com os professores Manoel Sant'Ana, Cassiano Rösing, Fabrício Collares, Deise Ponzoni, Fernando Hugo na Comissão de Pós-Graduação, com os quais pude compreender melhor a importância da Pós-Graduação e me motivei ainda mais para buscar a carreira acadêmica.

À Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), à Faculdade de Odontologia e à Universidade Federal do Rio Grande do Sul por tornar possível a realização de todas as pesquisas desenvolvidas durante a minha pós-graduação.

NOTA PRELIMINAR

A presente Tese foi redigida segundo a Resolução nº 093/2007 do Conselho de Ensino, Pesquisa e Extensão da Universidade Federal do Rio Grande do Sul. Enquadra-se na forma descrita no item "c" do artigo 3º da Resolução: "Tese, Dissertação ou Trabalho de Conclusão de Curso que contenham artigo(s) já publicado(s)".

RESUMO

O tratamento de canal ideal compreende a desinfecção e selamento do sistema de canais radiculares, de forma a impedir a reinfecção e permitir o reparo dos tecidos periapicais. Para isso é necessário o emprego de materiais que promovam um perfeito selamento, o que até o presente momento não foi completamente alcançado. O objetivo do presente estudo in vitro foi desenvolver um cimento endodôntico à base de resina de glicerol salicilato e α-fosfato tricálcico (αTCP), que apresente potencial de estimular a deposição mineral. Os cimentos foram formulados a partir de 70% em massa de uma resina de glicerol salicilato, obtida a partir da reação de transesterificação do salicilato de metila com o glicerol, e 30% em massa de carga inorgânica, composta de hidróxido de cálcio e αTCP, em diferentes proporções: 30% Ca(OH)₂, 25% Ca(OH)₂ + 5% αTCP, 20% Ca(OH)₂ + 10% α TCP, 15% Ca(OH)₂ + 15% α TCP. A resina sintetizada foi caracterizada por cromatografia de exclusão por tamanho, ressonância magnética de prótons e espectroscopia Raman. Os cimentos foram caracterizados quanto ao tempo de presa, degradação in vitro, potencial de alcalinização do meio, citotoxicidade e capacidade de estimular a deposição mineral. A resina de glicerol salicilato foi obtida por meio da reação de transesterificação, com 72% de rendimento. Os grupos avaliados apresentaram um tempo de presa de 240 min a 405 min e capacidade de alcalinizar o meio. Além disso, o aumento na concentração de αTCP diminuiu a degradação dos cimentos. A viabilidade celular após 48 h de contato direto com os cimentos experimentais não apresentou diferença quando comparada a um cimento obturador comercial (AH Plus) usado como referência. Os cimentos contendo 10% e 15% de αTCP exibiram a presença de uma camada superficial rica em fosfatos de cálcio após a imersão em água e simulated body fluid (SBF) por sete dias, verificada por espectroscopia Raman e MEV-EDS. Os resultados mostraram êxito no desenvolvimento do cimento endodôntico bioativo à base de resina de glicerol salicilato e α-fosfato tricálcico.

Palavras-chave: Cimentos dentários. Fosfatos de cálcio. Hidróxido de cálcio. Obturação do canal radicular.

ABSTRACT

Dental root canal treatment comprises the disinfection and complete filling of root canals in order to prevent the reinfection and allows the healing of periapical tissues. The use of materials that promotes an effective sealing has no achieved at the moment. The aim of this in vitro study was to develop a root canal sealer based on a glycerol salicylate resin and α -tricalcium phosphate (α TCP) that presents potential to stimulate mineral deposition. Sealers were formulated based on 70wt% of a glycerol salicylate resin, obtained by transesterification of methyl salicylate and glycerol, and 30wt% of inorganic filler, composed by calcium hydroxide and αTCP, in different proportions: 30% Ca(OH)₂, 25% Ca(OH)₂ + 5% α TCP, 20% Ca(OH)₂ + 10% α TCP, 15% Ca(OH)₂ + 15% αTCP. Synthesized resin was characterized by proton nuclear magnetic resonance spectroscopy, size exclusion chromatography and Raman spectroscopy. Sealers were characterized based on setting time, in vitro degradation over time, pH, cytotoxicity and mineral deposition. The glycerol salicylate resin was obtained by transesterification reaction, with 72% yield. Sealers presented setting time ranging from 240 min to 405 min, and basic pH over 8.21 after 28 days. Higher αTCP concentration leads to sealers with low solubility. Cell viability after 48 h in direct contact with sealers was similar to a commercial sealer (AH Plus) used as reference. The 10% and 15% αTCP sealers exhibited a calcium-phosphate layer on its surface after immersion in water and simulated body fluid (SBF) for 7 days, verified by Raman spectroscopy and MEV-EDX. The bioactive root canal sealer based on glycerol salicylate resin containing αTCP was successfully developed.

Key-words: Calcium Hydroxide. Calcium Phosphates. Dental Cements. Root Canal Obturation.

SUMÁRIO

1. INTRODUÇÃO	10
2. OBJETIVO	14
3. ARTIGOS	15
3.1. Artigo I	16
3.2. Artigo II	30
4 CONSIDERAÇÕES FINAIS	46
REFERÊNCIAS	48

1. INTRODUÇÃO

O preenchimento dos canais radiculares com materiais obturadores, incluindo-se os cimentos endodônticos e materiais de corpo, como a guta-percha, visa tornar inviáveis as bactérias residuais, aprisionando-as nos canalículos e túbulos dentinários, e evitar a percolação de fluidos via canal radicular, estando a qualidade da obturação relacionada ao sucesso da terapia (GILLEN et al., 2011; NG et al., 2011).

Até o momento, que se tenha conhecimento, nenhum cimento endodôntico disponível no mercado, associado a qualquer técnica obturadora já desenvolvida, é capaz de selar completamente o ápice radicular. Diversos tipos de cimentos, com diferentes composições químicas, foram propostos para serem usados em combinação com a guta-percha, com a finalidade de melhorar o selamento, entretanto, até o momento, sem alcançar um vedamento completo (YUCEL et al., 2006).

Cimentos obturadores contendo resina de salicilato (WALLER; JANDOUREK, biocompatibilidade, propriedade 1999) apresentam boa antimicrobiana desempenho clínico aceitável (LEONARDO et al., 1997; SILVA et al., 1997; BERBERT et al., 2002; WALTIMO et al., 2001; FARIA-JUNIOR et al., 2013), entretanto, apresentam elevada solubilidade (FARIA-JUNIOR et al., 2013), podendo levar a uma redução da longevidade da obturação. Visto isso, o desenvolvimento de uma resina de salicilato com estrutura química diferente das resinas de isobutil salicilato presente nos cimentos comerciais, poderia suplantar as limitações dos materiais disponíveis. As resinas de salicilato geralmente são uma blenda de polissalicilatos multifuncionais, sintetizadas a partir da transesterificação de ésteres, principalmente salicilato de metila, e alcoóis, como o neopentilglicol e o trimetilolpropano (WALLER; JANDOUREK, 1999). Uma alternativa inovadora seria a síntese da resina de salicilato a partir do glicerol, um álcool produzido em larga escala como subproduto da obtenção do biodiesel (SALVI; PANWAR, 2012). Para cada dez partes de biodiesel produzido, uma parte de glicerol bruto é gerada (KARINEN; KRAUSE, 2006; CHI et al., 2007), o que tem estimulado pesquisadores de todo mundo a procurar novos empregos ao glicerol. O glicerol, largamente

utilizado na indústria farmacêutica (FLUHR *et al.*, 2008) e de alimentos (HERNANDEZ-IZQUIERDO; KROCHTA, 2008) representa uma alternativa para a obtenção de resinas de salicilato, as quais podem ser utilizadas como matriz carreadora para íons ou outras substâncias na formulação de novos cimentos endodônticos. Ainda, o glicerol é um precursor para a síntese de triacilglicerol e fosfolipídeos no fígado e tecido adiposo, o que indica a biocompatibilidade do composto (MIGNECO *et al.* 2009).

Estudos demonstraram a viabilidade em se realizar a remineralização da superfície dentinária a partir da deposição mineral (KIM et al., 2010; Mai et al., 2010). Existem duas alternativas para desencadear esse tipo de remineralização, referidas na literatura como "bottom-up" e "top-down". Na primeira abordagem são utilizados análogos biomiméticos de proteínas da matriz, tais como os ácidos poliacrílico e polivinilfosfônico, que estabilizam os cristais de fosfato de cálcio nanoprecursores em núcleos de crescimento no interior da matriz de colágeno desmineralizada, de onde propaga-se a remineralização. O segundo método consiste em saturar o meio com íons cálcio e fosfato, de forma que os mesmos precipitem sobre a superfície a ser mineralizada. Em ambas as situações se faz necessária a presença de algum material solúvel que possa fornecer íons a um meio aquoso, sendo comumente utilizados cimentos de silicato de cálcio para esse fim (LIU et al., 2011). Alani e colaboradores em 2009, utilizando um compósito experimental à base de policaprolactona e um vidro fosfatado, em uma abordagem "top-down", mostraram ausência de infiltração, aferida por corante, em raízes obturadas com o referido material. Os autores relacionam o selamento obtido à presença de deposições minerais na interface do material obturador com a dentina radicular.

A utilização de materiais que estimulem a formação de tecido mineralizado (cemento/osso) na região periapical, após o tratamento endodôntico radical é desejada. Diversos biomateriais tem sido empregados para reparo de defeitos ósseos, destacando-se os cimentos de silicato de cálcio e os cimentos de fosfato de cálcio. Os materiais utilizados para reparo devem possuir um conjunto de propriedade físicas, químicas e biológicas que permitam desempenhar a função desejada e propiciar uma resposta adequada dos tecidos (GUASTALDI; APARECIDA, 2010). Os dentes e ossos dos vertebrados são um compósito natural formado por fibras colágenas envoltas por um sólido inorgânico (GUASTALDI;

APARECIDA, 2010). Sabe-se que apesar de não possuir composição estritamente definida e apresentar variações entre os estágios de maturação e envelhecimento, a estrutura cristalina e a composição da fase inorgânica dos tecidos duros dos vertebrados assemelha-se à hidroxiapatita, e por essa razão, os cimentos de fosfato de cálcio apresentam-se como os principais biomateriais utilizados para a reposição e regeneração de tecido ósseo (DOROZHKIN, 2010; DOROZHKIN, 2013).

Diversos cimentos de fosfato de cálcio tem sido utilizados como biomateriais, apresentando diferentes razões cálcio/fósforo, conforme o quadro 1. Um dos fatores que pode predizer o comportamento *in vivo* desses cimentos é a solubilidade dos fosfatos de cálcio em água, que de forma geral, é inversa a relação Ca/P do composto. O fosfato tricálcico, que pode ser sintetizado em diferentes formas alotrópicas, apresenta boa osteocondutividade, potencial de osteoindução e capacidade de estimular a proliferação celular. Sendo essas propriedades maiores para o αTCP em comparação com o βTCP, provavelmente devido à sua estrutura cristalina, que possibilita uma maior troca de íons com o meio, além de sua maior solubilidade em pH fisiológico (GUASTALDI; APARECIDA, 2010; GANDOLFI *et al.*, 2010; LI *et al.*, 2012). Ainda, como resultado da reação do αTCP com água, há a precipitação de hidroxiapatita (GINEBRA *et al.*, 1997; MOREJON-ALONSO, *et al.*, 2012).

Quadro 1. Tipos de fosfatos de cálcio e suas solubilidades

Razão molar Ca/P	Nome e abreviação usual	Fórmula química	Solubilidade em água a 25°C (g/l)
0,5	Fosfato monocálcico (MCP)	Ca(H ₂ PO ₄) ₂	~17
1,0	Mono-hidrogênio fosfato de cálcio (DCP)	CaHPO₄	~0,048
1,33	Fosfato octacálcico (OCP)	Ca ₈ (HPO ₄) ₂ (PO ₄) ₄ .5H ₂ O	~0,0081
1,5	α-fosfato tricálcico (αTCP)	α -Ca ₃ (PO ₄) ₂	~0,0025
1,5	β-fosfato tricálcico (βTCP)	β -Ca ₃ (PO ₄) ₂	~0,0005
1,2 - 2,2	Fosfatos de cálcio amorfo (ACP)	$Ca_xH_y(PO_4)_z$. nH_2O	*
1,67	Hidroxiapatita (HÁ)	Ca ₁₀ (PO ₄) ₆ (OH) ₂	~0,0003
1,67	Fluorapatita (FA)	Ca ₁₀ (PO ₄) ₆ F ₂	~0,0002
2,0	Fosfato tetracálcico (TTCP)	Ca ₄ (PO ₄) ₂ O	~0,0007

Adaptado de Dorozhkin, 2013. * não pode ser determinada precisamente.

Sendo assim, o desenvolvimento de um cimento à base de resina de glicerol salicilato e α-fosfato tricálcico, que propicie as condições necessárias à precipitação mineral na interface obturação/dentina e o reparo dos tecidos periapicais, apresentase como uma alternativa promissora para otimização dos tratamentos endodônticos.

2. OBJETIVO

O objetivo do presente estudo foi desenvolver um cimento endodôntico à base de resina de glicerol salicilato e α-fosfato tricálcico, que apresente potencial de estimular a deposição mineral.

Dessa forma, em um primeiro momento objetivou-se:

- Sintetizar uma resina de glicerol salicilato;
- Caracterizar as propriedades da resina obtida e verificar a aplicabilidade para o uso em um cimento endodôntico.

A resina sintetizada foi utilizada em cimentos "modelo" contendo diferentes concentrações de α-fosfato tricálcico. Nesse momento objetivou-se caracterizar os cimentos quanto às:

- Propriedades químicas e físicas;
- Citotoxicidade;
- Capacidade de promover a deposição de fosfatos de cálcio.

3. ARTIGOS

O corpo da presente tese é composto por dois artigos:

- Artigo I: Synthesis and characterization of a glycerol salicylate resin for bioactive root canal sealers. Publicado no periódico International Endodontic Journal (doi: 10.1111/iej.12149) (PORTELLA et al., 2014);
- Artigo II: Glycerol salicylate-based containing α-tricalcium phosphate as a bioactive root canal sealer. Publicado no periódico Journal of Biomedical Materials Research Part B: Applied Biomaterials (doi: 10.1002/jbm.b.33326) (PORTELLA et al., 2015).

Os manuscritos, formatados de acordo com os requisitos dos periódicos aos quais foram submetidos, encontram-se a seguir.

3.1. Artigo I

Synthesis and characterization of a glycerol salicylate resin for bioactive root canal sealers

F F Portella¹

P D Santos¹

G B Lima²

V C B Leitune¹

C L Petzhold²

F M Collares^{1*}

S M W Samuel¹

¹Laboratory of Dental Materials, School of Dentistry ²Institute of Chemistry, Federal University of Rio Grande do Sul, Porto Alegre, RS, Brazil

Running title: Glycerol salicylate resin

Key-words: salicylate resin, salicylate, glycerol, bioactive sealers, RAMAN.

*Corresponding Author:

Fabrício Mezzomo Collares

Laboratory of Dental Materials, School of Dentistry, Federal University of Rio Grande do Sul, Rua Ramiro Barcelos 2492, Porto Alegre, RS, Brazil, 90035-003

Telephone: 55 51 33085198

Email: fabricio.collares@ufrgs.br

Abstract

Aim To develop and characterize a salicylate resin with potential use in bioactive endodontic sealers.

Methodology Methyl salicylate, glycerol, and titanium isopropoxide were added in a closed system for the transesterification reaction. The resin obtained was characterized by proton nuclear magnetic resonance spectroscopy (1H NMR) and size exclusion chromatography (SEC). To verify the applicability of the resin to the development of endodontic sealers, experimental cements were prepared by mixing glycerol salicylate resin, calcium hydroxide, and methyl salicylate in ratios of 2:1:1, 1:2:1, 1:1:2, 1:1:1, 4:1:1, 1:4:1, and 1:1:4. Setting times were measured according to ISO 6876. Features of the hardening reaction were described by micro-RAMAN spectroscopy.

Results The transesterification reaction had a 72% efficiency. The 1H NMR analysis revealed the presence of the expected functional groups (hydroxyls and aromatic rings), and the SEC confirmed the molar mass of the resin produced. The setting times of experimental sealers ranged from 70 min (ratio 1:1:1) to 490 min (ratio 1:1:4). The conversion of the salicylic groups (1 613 cm-1) to salicylate salt (1 543 cm-1) and the reduction of calcium hydroxide peaks (1084 cm-1 and 682 cm-1) were confirmed by micro-RAMAN spectroscopy, which showed the calcium chelation by the resin.

Conclusion The new glycerol salicylate resin was successfully synthesized and revealed a potential application in the development of endodontic sealers.

Introduction

The main objective of root canal treatment is to heal and/or to prevent the contamination of periapical tissues. Filling of the canal system plays an important role in achieving this outcome, and the materials used as root canal sealers should not be cytotoxic. Additionally, they ideally should be biological stimulators, as the cements and bulk materials are in contact with dentine and periapical tissues (Leonardo *et al.* 1995, Tanomaru Filho *et al.* 1998). Therefore, root canal sealers that have ionic delivery have been proposed to enhance sealing of the apical region by mineral deposition to the canal walls (Alani *et al.* 2009).

Salicylate resin was introduced as a resin matrix for endodontic sealers in the early nineties (Waller & Jandourek 1999). Commercial sealers that contain salicylate resin have presented reliable biocompatibility (Leonardo *et al.* 1997, Silva *et al.* 1997), satisfactory antibacterial properties (Berbert *et al.* 2002), and acceptable clinical performances (Waltimo *et al.* 2001). However, commercial salicylate based sealers present high solubility (Faria-Júnior *et al.* 2013) leading to decreased treatment longevity. Synthesis of novel salicylate sealers with improved properties are required to overcome these limitations.

Resins based on salicylate are a blend of multifunctional polysalicylates, synthesized by the transesterification of esters (mainly methyl salicylate) and alcohols, such as neopentylglycol and trimethylolpropane (Waller & Jandourek 1999). An innovative alternative to salicylate resins is the use of glycerol, an alcohol that is produced on a large scale as a biodiesel byproduct (Salvi & Panwar 2012). For every ten parts of biodiesel produced, one part of crude glycerol is generated as a byproduct (Karinen & Krause 2006, Chi *et al.* 2007). Glycerol, widely used in the pharmaceutical (Fluhr *et al.* 2008) and food industries (Hernandez-Izquierdo & Krochta 2008), is an alternative to the production of salicylate resins, which can serve as a carrying matrix for ions or other substances in the formulation of endodontic sealers. Thus, the purpose of this study was to develop and characterize a salicylate resin to be used in bioactive endodontic sealers.

Material and Methods

Resin synthesis

The salicylate resin was synthesized by transesterification of glycerol (Vetec Química Fina Ltda, Duque de Caxias, RJ, Brazil, 99.5%) and methyl salicylate (Vetec

Química Fina Ltda, Duque de Caxias, RJ, Brazil, 99%). The reagents were added in a ratio of 1:7.5 (w/w) in a closed vacuum (10⁻² mmHg) system (a volumetric flask coupled to a vacuum pump and a condenser to simple distillation) and heated at 100°C for 1 hour. The vacuum was removed, the titanium isopropoxide (Sigma-Aldrich, St. Louis, MO, USA, 97%) was added as a catalyst agent (0.05% w/w) at 60°C, and the temperature of the system was increased to 204°C for 17 hours. The temperature was decreased to 178°C until methanol, obtained as byproduct, was eliminated by simple distillation. The vacuum was reapplied to remove the excess methyl salicylate. The distillation of methanol and vacuum application were used to purify the resin. Figure 1 shows the molecular structures of reagents and the expected products of the transesterification reaction.

Figure 1. Molecular structure of (a) glycerol, (b) methyl salicylate, (c) glycerol monosalicylate (MW=212.20), (d) glycerol disalicylate (MW=332.30), and (e) glycerol trisalicylate (MW=452.41).

Characterization of the resin

The molecular weights of the products were determined with a size exclusion chromatography (SEC) composed of a tetra detector array (TDA 302, VISCOTEK VE2001, Houston, TX, USA) and a polystyrene column, THF was used as solvent with a flow rate of 1mL/min. The molecular structure was confirmed by proton nuclear

magnetic resonance spectroscopy (¹H NMR) with a VARIAN VNMRS 300 instrument (Palo Alto, CA, USA), using deuterated chloroform as a solvent and tetramethylsilane as an internal standard.

Experimental cements

Experimental cements were formulated and tested to confirm that the resin obtained was applicable to the development of root canal sealers. The applicability was verified by mixing resin, calcium hydroxide (Biodinâmica, Ibiporã, PR, Brazil, p.a.) and methyl salicylate in different proportions by weight: 1:1:1, 2:1:1, 1:2:1, 1:1:2, 4:1:1, and 1:1:4. Hardening and setting times of experimental cements were verified according to ISO 6876 (2001). The cements were manipulated and placed on cylindrical matrixes, 10 mm in diameter and 2 mm in height. After 120 seconds of mixing, indentations were formed on the surface of the specimens with a 100 g \pm 0.5 g cylindrical indenter (Ø2 \pm 0.1 mm). Indentations were made until no indentations could be visualized by operator naked eye, and the time of this occurrence was registered as the setting time. The test was repeated 3 times, and the setting time was calculated as the mean of the measures.

The chemical reaction between the resin and calcium hydroxide was characterized by micro-Raman spectroscopy (Senterra, Bruker Optics, Ettlingen, Germany). Figures 2 and 3 show the Raman spectrum of each reagent separately. The cement (proportion 1:1:1) was prepared, and the surface was excited by a 785 nm laser, with 100 mW of power, 10 seconds of integration time, and 2 co-additions. One spectrum was obtained immediately after mixing and after the setting time. The spectrums were submitted to a concave rubberband correction with one interaction and 64 baseline points on Opus 6.5 software (Bruker Optics, Ettlingen, Germany) and then the data was transferred to Microsoft Office Excel 2007 (Microsoft, Redmond, WA) to graph plotting. The chemical reaction was verified in a manner similar to that in a previous study (Mazinis *et al.* 2007).

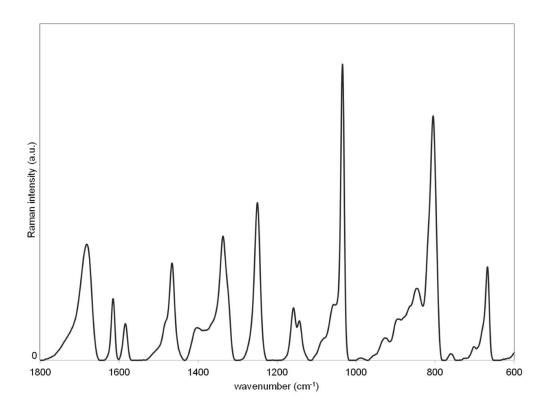


Figure 2. Raman spectrum of the glycerol salicylate resin.

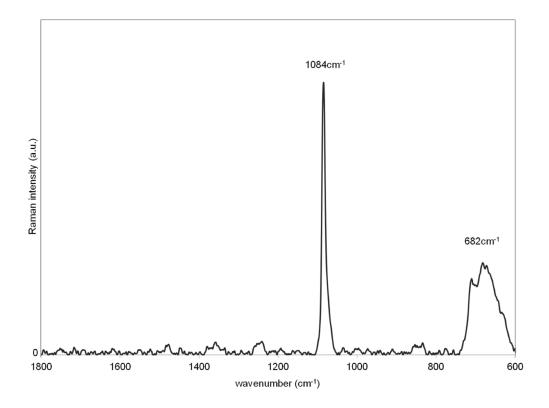


Figure 3. Raman spectrum of the commercial calcium hydroxide used to formulate the experimental cements.

Results

The yield of the transesterification reaction was 72%. The ¹H NMR analyses (Figure 4) revealed the presence of the expected proton signals of the functional groups for the proposed reaction: phenolic OH-protons appears at 10.5 ppm, the aromatic protons between 7 – 8 ppm, the methinic proton (-CH) of the glycerol moiety at 5.8 ppm and the methylenic protons (-CH₂) in the range of 4 – 5 ppm. The average molecular weight obtained from SEC (265 g/mol) indicated that the resin obtained composed of a mixture of 50-60% glycerol monosalicylate and 40-50% glycerol disalicylate, as this proportion, by weight, present a molecular weight range of 260.00 to 272.25g/mol. Only traces of glycerol trisalicylate were expected, since the central hydroxyl of the triol is less reactive than the extremities hydroxyls.

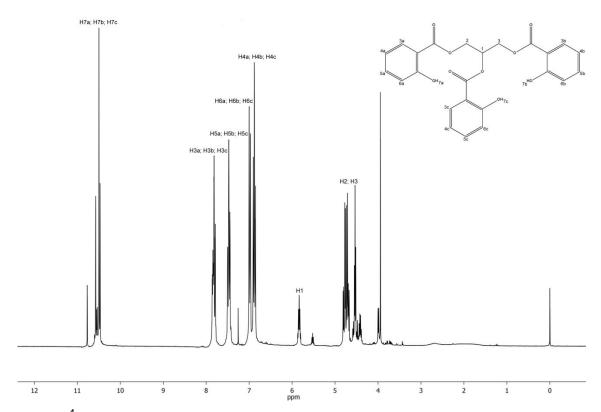


Figure 4. ¹H NMR spectrum of the resin and respective hydrogen assignments.

The setting time of the experimental cements ranged from 70 min for the 1:1:1 proportion to 490 min for the 1:1:4 proportion, as shown in Table 1. A 50% addition of methyl salicylate to the mixture produced an increase of the setting time, reaching 490 min.

Table 1. Setting time of experimental sealers as function of components proportion (in weigth)

resin:Ca(OH) ₂ :methyl	setting time (min)
salicylate	
1:1:1	70
2:1:1	135
1:2:1	*
1:1:2	95
4:1:1	360
1:1:4	490

^{*}without handling condition

Micro-Raman spectroscopy (Figure 5) revealed the chelation of calcium by the hydroxyl resin salicylate groups, promoting the hardening of cements. The spectrum peaks of calcium hydroxide (1084 cm⁻¹ and 682 cm⁻¹ (Chaix-Pluchery et al. 1984)) decreased and a group of three absorptions was observed for the region between 1520 and 1640 cm⁻¹ which is characteristic of a structure that features the ester carbonyl group of the salicylate ligand and calcium-coordinated phenoxide as shown in Figure 6 (Dain et al. 2011). The highest frequency stretch (at 1613 cm⁻¹) corresponds to a vibrational mode that involves C-C stretching of the aromatic ring coupled to stretching of the phenoxy and carboxylic acid C-O groups. A second absorption, observed at 1576 cm⁻¹ is focused on C=O stretching of the ester carbonyl group, with coupling to an acid C-O-H wag and ring C-C stretching. The third (1543) cm⁻¹) corresponds to C_{ring}-C_{acid}-O stretching coupled to ring C-C stretches. A fourth prominent absorption is observed at 1153 cm⁻¹ corresponding to acid C-O-H wag coupled to ring C-C stretches. The Raman spectrum of methyl salicylate non-metal coordinated showed only two bands in the region around 1600cm⁻¹ (Varghese et al. 2007) as the glycerol salicylate resin (Figure 2).

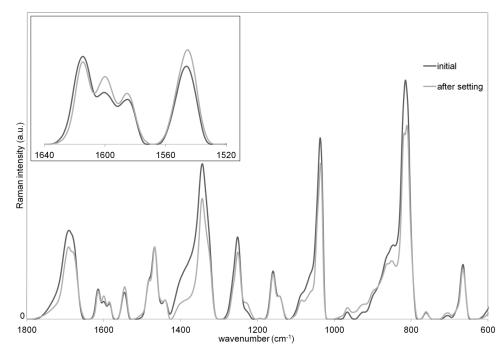


Figure 5. Raman spectrum of the setting reaction of the experimental sealer. The region of salicylic group peaks was enlarged to show the production of the salicylate salt.

Figure 6. Representative scheme of a structure that features a neutral carboxylic acid group of the salicylate ligand and calcium-coordinated phenoxide.

Discussion

Root canal treatment is comprised of the cleaning and shaping of the root canals by following the filling of the canal system. However, the sealing of the root canal system is still a concern in endodontics (Kopper *et al.* 2006, Kim *et al.* 2010). The use of bioactive materials to fill the root canal system could be a viable method to achieve effective sealing, obliterating the gaps between the filling material and the root canal walls, sealing dentinal tubules and forming a mineralized barrier at the

apex, as these materials could interact with periapical tissues (Collares *et al.* 2012). Alani *et al.* (2009) proposed a polycaprolactone—iron phosphate glass composite as a root canal filling material, which releases ions and achieves an appropriate seal via mineral precipitation on the filling/dentin interface, completely preventing fluid percolation through the root canal and filling material. Bioactive materials should be composed of active components, such as drugs and inorganic compounds, along with a carrier matrix with high biocompatibility and a controlled release of the active components. These biomaterials raise the possibility of potentially controlling the relative benefits of ion release to induce antibacterial effects and to regenerate periapical tissues (Gandolfi *et al.* 2011a, Gandolfi *et al.* 2011b). In this study, a glycerol salicylate resin was *synthesized* that could be adequate as a bioactive sealer.

The chemical groups noted during the NMR analysis and the molecular weight of the product obtained on SEC confirmed the presence of glycerol salicylate, which was the expected product of the transesterification reaction of glycerol and methyl salicylate. The applicability of the salicylate resin was verified by the formulation of experimental sealers that contained different concentrations of calcium hydroxide and methyl salicylate. Methyl salicylate was used to adjust the rheological properties of sealers and to reduce the viscosity of the glycerol salicylate resin. Setting times were registered; all formulations presented setting times, according to ISO 6876, except for the formulation containing 50% calcium hydroxide. This formulation did not present handling characteristics (impossible to achieve a homogeneous mixture), most likely due to a high filler concentration.

The hardening of cements occurs through calcium chelation by the hydroxyl of the salicylate groups of the resin, as characterized by micro-RAMAN spectroscopy (Figure 5). This correlates with the results of a previous study, in which this interaction was demonstrated by Fourier Transformed Infrared Spectroscopy (Mazinis *et al.* 2007). The groups with low concentrations of calcium hydroxide showed longer setting times. These results could be due to the presence of excess salicylate groups, which cause the failure of reaction stoichiometry. Theoretically, each mol of calcium hydroxide can react with 2 mol of the salicylate group. The molecular weight-average of the resin obtained on SEC was 265g/mol, indicating that glycerol monosalicylate (MW=212.20 g/mol) and glycerol disalicylate (MW=332.30 g/mol) are most likely the major components. Therefore, in a blend composed of 50-

50 mono and disalicylate, for each one part by mass of calcium hydroxide, approximately five parts by mass of resin will be necessary to fit the stoichiometric reaction.

The reaction could generate different products, according to the number of hydroxyl groups of glycerol that participates in the reaction. The resin obtained could be a mixture of mono-, di-, or trisalicylate glycerol, presenting molecular weights of 212.20, 322.30, and 452.41g/mol, respectively. The major yield of the reaction will be achieved when one glycerol molecule reacts with three methyl salicylate molecules; for this to take place, for each part of glycerol, five parts, in mass, of methyl salicylate will be necessary. Methyl salicylate was added in excess of 50% to ensure that the entire amount of glycerol reacted. This feature of the synthesis process could explain its 72% efficiency because part of the total mass of reagents was converted to methanol (byproduct) and probably some methyl salicylate remained unreacted.

The characteristics of synthesized salicylate resin could be used as a promising carrier agent for the development of bioactive endodontic sealers. Glycerol has excellent biocompatibility (it is a precursor for the synthesis of triacylglycerols and of phospholipids in the liver and adipose tissue) (Migneco *et al.* 2009) that could be used in calcium based sealers to promote remineralization when in contact with connective tissues. Moreover, salicylates are anti-inflammatory agents (Duthie & Wood 2011) that are widely used in the pharmacological industry (Amann & Peskar 2002). To combine the outstanding properties of glycerol and salicylate in a resin seems to be a promising approach in the development of endodontic sealers, especially if those materials are in contact with periapical tissues.

Conclusion

The salicylate resin synthesized in this study was successfully generated, and it exhibited applicability in bioactive endodontic sealers that contained inorganic components, that permit its incorporation into the matrix resin by chelation.

Acknowledgments

The authors gratefully acknowledge CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) and CNPq (Conselho Nacional de Pesquisa) for the scholarship (F.F.P. and V.C.B.L; and P.D.S).

References

- Alani A, Knowles JC, Chrzanowski W, Ng YL, Gulabivala K (2009) Ion release characteristics, precipitate formation and sealing ability of a phosphate glass-polycaprolactone-based composite for use as a root canal obturation material. *Dental Materials* **25**, 400-10.
- Amann R, Peskar BA (2002) Anti-inflammatory effects of aspirin and sodium salicylate. *European Journal of Pharmacology* **447**, 1-9.
- Berbert FL, Leonardo MR, Silva LA, Tanomaru Filho M, Bramante CM (2002) Influence of root canal dressings and sealers on repair of apical periodontitis after endodontic treatment. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology & Endodontics* **93**, 184-9.
- Chaix-Pluchery O, Ciosmak D, Niepce JC, Peyrard M (1984) Raman study of prereactional transformations in calcium hydroxide crystals during a thermal treatment leading to dehydration. *Journal of Solid State Chemistry* **53**, 273-6.
- Chi ZY, Pyle D, Wen ZY, Frear C, Chen SL (2007) A laboratory study of producing docosahexaenoic acid from biodiesel-waste glycerol by microalgal fermentation. *Process Biochemistry* **42**, 1537-45.
- Collares FM, Leitune VC, Rostirolla FV, Trommer RM, Bergmann CP, Samuel SM (2012) Nanostructured hydroxyapatite as filler for methacrylate-based root canal sealers. *International Endodontic Journal* **45**, 63-7.
- Dain RP, Gresham G, Groenewold GS, Steill JD, Oomens J, van Stipdonk MJ (2011) Infrared multiple-photon dissociation spectroscopy of group II metal complexes with salicylate. *Rapid Communications in Mass Spectrometry* **25**, 1837–46.
- Duthie GG, Wood AD (2011) Natural salicylates: foods, functions and disease prevention. *Food Funct* **2**, 515-20.
- Faria-Júnior NB, Tanomaru-Filho M, Berbert FL, Guerreiro-Tanomaru JM (2013)

 Antibiofilm activity, pH and solubility of endodontic sealers. *International Endodontic Journal*. in press.
- Fluhr JW, Darlenski R, Surber C (2008) Glycerol and the skin: holistic approach to its origin and functions. *British Journal of Dermatology* **159**, 23-34.
- Gandolfi MG, Shah SN, Feng R, Prati C, Akintoye SO (2011)a Biomimetic calciumsilicate cements support differentiation of human orofacial mesenchymal stem cells. *Journal of Endodontics* **37**, 1102-8.

- Gandolfi MG, Taddei P, Siboni F, Modena E, De Stefano ED, Prati C (2011)b Biomimetic remineralization of human dentin using promising innovative calcium-silicate hybrid "smart" materials. *Dental Materials* **27**, 1055-69.
- Hernandez-Izquierdo VM, Krochta JM (2008) Thermoplastic processing of proteins for film formation--a review. *Journal of Food Science* **73**, R30-9.
- ISO 6876 (2001) Dental root canal sealing materials, 2nd edn. Geneva, Switzerland: International Organization for Standardization.
- Karinen RS, Krause AOI (2006) New biocomponents from glycerol. *Applied Catalysis A: General* **306,** 128-33.
- Kim YK, Grandini S, Ames JM *et al.* (2010) Critical review on methacrylate resinbased root canal sealers. *Journal of Endodontics* **36**, 383-99.
- Kopper PM, Vanni JR, Della Bona A, de Figueiredo JA, Porto S (2006) In vivo evaluation of the sealing ability of two endodontic sealers in root canals exposed to the oral environment for 45 and 90 days. *Journal of Applied Oral Science* **14,** 43-8.
- Leonardo MR, Almeida WA, da Silva LA, Utrilla LS (1995) Histopathological observations of periapical repair in teeth with radiolucent areas submitted to two different methods of root canal treatment. *Journal of Endodontics* **21**, 137-41.
- Leonardo MR, Silva LA, Utrilla LS, Assed S, Ether SS (1997) Calcium hydroxide root canal sealers--histopathologic evaluation of apical and periapical repair after endodontic treatment. *Journal of Endodontics* **23**, 428-32.
- Mazinis E, Eliades G, Lambrianides T (2007) An FTIR study of the setting reaction of various endodontic sealers. *Journal of Endodontics* **33**, 616-20.
- Migneco F, Huang YC, Birla RK, Hollister SJ (2009) Poly(glycerol-dodecanoate), a biodegradable polyester for medical devices and tissue engineering scaffolds. *Biomaterials* **30**, 6479-84.
- Salvi BL, Panwar NL (2012) Biodiesel resources and production technologies A review. *Renewable and Sustainable Energy Reviews* **16,** 3680-9.
- Silva LA, Leonardo MR, Faccioli LH, Figueiredo F (1997) Inflammatory response to calcium hydroxide based root canal sealers. *Journal of Endodontics* **23**, 86-90.
- Tanomaru Filho M, Leonardo MR, Silva LA, Utrilla LS (1998) Effect of different root canal sealers on periapical repair of teeth with chronic periadicular periodontitis. *International Endodontic Journal* **31**, 85-9.

- Varghese HT, Yohannan Panicker C, Philip D, Mannekutla JR, Inamdar SR (2007) IR, Raman and SERS studies of methyl salicylate. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy* **66,** 959-63.
- Waller DE, Jandourek E, Inventors; The Kerr Corporation, assignee (1999) Dental Resins. United States Patent no. 5922785.
- Waltimo TM, Boiesen J, Eriksen HM, Orstavik D (2001) Clinical performance of 3 endodontic sealers. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology & Endodontics* **92,** 89-92.

3.2. Artigo II

Glycerol salicylate-based containing α -tricalcium phosphate as a bioactive root canal sealer

Fernando F. Portella¹

Fabrício M. Collares^{1*}

Luís A. dos Santos²

Bruno P. dos Santos³

Melissa Camassola³

Vicente C. B. Leitune¹

Susana M. W. Samuel¹

¹Dental Materials Laboratory, School of Dentistry, Federal University of Rio Grande do Sul, Porto Alegre, RS, Brazil

²Biomaterials Laboratory, Department of Materials, Federal University of Rio Grande do Sul, Porto Alegre, RS, Brazil

³Laboratory of Stem Cells and Tissue Engineering, Universidade Luterana do Brasil, Canoas, RS, Brazil

*Corresponding Author:

Fabrício Mezzomo Collares

Dental Materials Laboratory, School of Dentistry, Federal University of Rio Grande do Sul, Rua Ramiro Barcelos 2492, Porto Alegre, RS, Brazil, 90035-003

Telephone: 55 51 33085198

Email: fabricio.collares@ufrgs.br

ABSTRACT

The use of bioactive materials instead of inert materials to fill the root canal space could be an effective approach to achieve a hermetic seal and stimulate the healing of periapical tissues. The purpose of this study was to develop and characterize an endodontic sealer based on a glycerol salicylate resin and α -tricalcium phosphate (α TCP) at physical and chemical properties. Different sealers were formulated using 70% of a glycerol salicylate resin and 30% of a mixture of calcium hydroxide and α TCP (0, 5, 10 or 15%, in weight). Sealers formulated were characterized based on setting time, *in vitro* degradation over time, pH, cytotoxicity and mineral deposition. Sealers presented setting time ranging from 240 to 405 min, and basic pH over 8.21 after 28 days. Higher α TCP concentration leads to sealers with low solubility. Cell viability after 48 h in direct contact with sealers was similar to a commercial sealer used as reference. The 10% and 15% α TCP sealers exhibited a calcium-phosphate layer on the surface after immersion in water and SBF for 7 days. Glycerol salicylate sealers with 10% and 15% α -tricalcium phosphate showed reliable physical chemical properties and apatite-forming ability.

Keywords: α-tricalcium phosphate, bioactive sealers, glycerol, salicylate, Raman.

Running Head: α*TCP bioactive root canal sealer*

INTRODUCTION

Dental root canal treatment comprises the disinfection and complete filling of root canals in order to heal the periapical tissues. The quality of sealing promoted by endodontic sealers is one of the main reasons for clinical success¹. Recently, resinbased sealers (e.g., epoxy, methacrylate and salicylate based) were introduced for endodontic treatment to improve sealing and resistance to degradation².

The salicylate resins present a reliable clinical performance³ due to the antiinflammatory⁴ and biocompatibility⁵ properties with a high solubility observed⁶.

Portella *et al.*⁷ synthesized a trifunctional salicylate resin with 72% efficiency in
transesterification reactions at a clinically acceptable setting time. To increase
mechanical properties of root canal sealers, bioactive inorganic fillers are
incorporated in a resin matrix, such as calcium phosphates. Calcium phosphatebased sealers induce a favorable cell response⁸, release of calcium and hydroxyl
ions⁹ and stimulated deposition of apatite¹⁰. However, commercial available calcium
phosphate-based sealers present high solubility, decreasing the clinical success of
root canal treatment¹¹.

In this way, combining glycerol salicylate resin and α TCP seems to be a promising approach to increase the quality of endodontic sealers. The purpose of this study was to evaluate the influence of α -tricalcium phosphate (α TCP) addition to a glycerol salicylate resin at physical and chemical properties.

MATERIAL AND METHODS

Experimental sealers were obtained by mixing a glycerol salicylate resin, synthesized according to Portella *et al.*⁷, calcium hydroxide and α -tricalcium phosphate. All sealers were composed by 70wt% resin and 30wt% inorganic components. The inorganic portion was a mixture of calcium hydroxide and α TCP. The content of α TCP used was 0, 5, 10 and 15 wt% in designed experimental groups. The calcium hydroxide (Ca(OH)₂ p.a., Biodinâmica, Ibiporã, PR, Brazil, batch no. 86812) and α TCP, synthesized according to Santos *et al.*¹², had a particle size analyzed by laser diffraction analyzer (CILAS 1180 Particle-Size-Analyzer, Compagnie Industrielle des Lasers, Orleans, Loiret, France). The reagents were hand mixed, inserted in a silicon matrix and covered by polyester film to obtain samples for each of the following tests.

Setting time

Time required to harden the sealers was assessed according to a previous study⁷. The sealers were manipulated, placed on cylindrical matrixes and stored in a humid cabinet at 36° C. Indentations were formed on the surface of the specimens with a 100-g and 2-mm diameter needle, as specified on ISO 6876¹³, until no indentations could be visualized by the operator's naked eye, and the time of this occurrence, it was registered as the setting time. The setting time was calculated as the mean of three measures.

In vitro degradation and pH analysis

The weight loss of experimental sealers was determined after 24 hours, 7 days and 28 days of storage in water and in simulated body fluid (SBF). The SBF preparation was described elsewhere ¹⁴. Three discs 10±0.1 mm in diameter and 1.0±0.1 mm thick were immersed in 10 ml of distillated water or SBF and stored at 36 °C for each period. The dried mass of specimens was determined before and after the immersion. To obtain the dried mass, the specimens were placed in desiccators containing silica gel and stored at 36 °C. Each specimen was weighed repeatedly at 24-h intervals until a constant mass was obtained (until the specimen mass's decrease did not differ more than 0.1 mg in a 24-h period). The weight loss (WL) percentage was calculated according to the following equation ¹⁵:

$$WL (\%) = (W_f W_o)/W_o \times 100$$

Where W_f is the weight after immersion and W_0 is the weight before immersion. The mean weight loss of three specimens was recorded for each experimental sealer. Two-way ANOVA and Tukey's test were used to assess differences between groups submitted to the same storage solution. Comparisons were made at 5% significance.

The pH of the storage water and SBF were evaluated using a digital pHmeter (pHmetro DM-23, Digicrom Analytical Ltda, São Paulo, SP, Brazil). Three measurements were performed for each period: 24 hours, 7 days and 28 days. Changes in pH were assessed by two-way ANOVA followed by Tukey's multiple comparison test.

Bioactivity assay: Raman and SEM-EDX surface characterization

Glycerol salicylate resin and inorganic reagents were mixed, inserted in a cylindrical matrix, and covered by a polyester coverslip to form disks 6.5 ± 0.1 mm in diameter and 1.5 ± 0.1 mm thick. After preparation, samples were stored in a humid cabinet until hardening. Three disks of each experimental sealer were produced and the surfaces were characterized by Raman spectroscopy and by SEM-EDX immediately after hardening and after storage in 10 ml of distillated water or SBF for 7 days at $36 \, ^{\circ}\text{C}^{14}$. Before analysis the groups submitted to immersion were washed with 10 ml of distillated water for 20 s and stored in a desiccator. The SBF used had the following ion concentration, in mol/m³: Na⁺ (213.0), K⁺ (7.5), Mg²⁺ (2.3), Ca²⁺ (3.8), Cl⁻ (221.7), HCO³- (6.3), HPO₄³- (1.5) and SO₄²- (0.8). The pH was adjusted to 7.4.

A 200x200-µm area of the specimen's surface was analyzed by Raman spectroscopy (Senterra, Bruker Optics, Ettlingen, Germany). The sample surface was irradiated by a 785-nm laser at 50 mW on 100 points equally distributed. Each point was irradiated 3 times for 5 s. Spectra were obtained at the region between 1800 and 440 cm⁻¹ in a backscattering mode. The integral peaks, 962 cm⁻¹ and 1613 cm⁻¹, corresponding to the symmetric stretching v_1 mode of phosphate¹⁶ and vibration mode of carbon-to-carbon stretching of the aromatic ring of salicylic group¹⁷, respectively, were calculated using Opus software (Opus 7.5, Bruker Optics, Ettlingen, Germany). To standardize the integral calculation peak areas, the entire region below peaks from 980 to 949 cm⁻¹ and 1624 to 1605 cm⁻¹ were considered. Changes on the ratio between the integral intensity of 962 cm⁻¹ and 1613 cm⁻¹ bands (A_{962cm}⁻¹/A_{1613cm}⁻¹) were used to analyze the formation of a apatite-like layer on the sealer surface after immersion in distillated water and SBF¹⁶.

The sample was carbon-sputtered and examined using scanning electron microscopy (JSM 5800, Jeol Ltd, Tokyo Japan) at 10 kV, followed by energy dispersive X-ray analysis to assess the elemental composition.

Cytotoxicity test

Cell viability was assessed by direct contact test using fibroblast cells (L929) (BCRJ batch no. 000604, Rio de Janeiro, Brazil) according to procedures described in ISO 10993-5¹⁸. The specimens used were 3.0±0.1 mm in diameter and 1.0±0.1 mm thick and were previously sterilized in hydrogen peroxide plasma. The MTT test

was performed in triplicate and the cell viability was read at 540 nm absorbance (Multiskan EX Microplate Reader, MTX Lab Systems Inc, VA, USA) after 24 or 48 h in direct contact with the sealers. Cells without any sealer were considered to be controlled for cytotoxicity, and samples of a widely used commercial sealer (AH Plus, Dentsply DeTrey GmbH, Konstanz, Germany) were used for comparison of the results. Data were transformed to log scale and analyzed by two-way ANOVA and Tukey's test.

RESULTS

Mean particle size of calcium hydroxide and α -tricalcium phosphate was 6.77 μ m (D_{10%}=1.24 μ m, D_{50%}=5.87 μ m and D_{90%}=13.75 μ m) and 6.03 μm $(D_{10\%}=1.03)$ μm, $D_{50\%} = 5.43$ μm and D_{90%}=12.00 µm), respectively. Setting time of sealers and changes in pH of SBF of the degradation test are shown in Table 1. Setting time ranged from 240 (0 α TCP) to 405 min (15%) αTCP). The pH of water turns basic in all groups after 24 h, the highest value (11.42±0.03) measured for 0 αTCP at 7 days. All sealers were able to increase the pH of SBF over time, reaching 8.94 ± 0.01 for 0 α TCP and 8.21 ± 0.04 for 15% αTCP groups, the highest and lowest pH after 28 days, respectively.

Immersion in distillated water and SBF reveals that all sealers had mass loss after soaking that increased with time, as shown in Figure 1. The sealers containing 10% and 15% α TCP showed less weight loss after 28 days of water or SBF immersion (p<0.05). Sealers without α TCP ranged from 72.49% to 60.63%. Groups with 15% α TCP showed a mass loss of 41.15% and 31.86% in water and SBF immersion, respectively.

Table 1. Setting time and pH of water and SBF according to experimental sealers.

Experimental Setting time	Setting time		Water pH			SBF pH	
sealer	(min)	24h	7 days 28 days	28 days	24h	24h 7 days 28 days	28 days
0 aTCP	240	7.62 (0.10) ^{Ac}	7.62 (0.10) ^{Ac} 11.42 (0.03) ^{Aa} 9.72 (0.03) ^{Ab} 7.68 (0.02) ^{Ac} 7.94 (0.05) ^{Ab} 8.94 (0.01) ^{Aa}	9.72 (0.03) ^{Ab}	7.68 (0.02) ^{Ac}	7.94 (0.05) ^{Ab}	8.94 (0.01) ^{Aa}
5 % αTCP	240	7.53 (0.01) ^{Ac}	7.53 (0.01) ^{Ac} 7.99 (0.18) ^{Ba} 9.78 (0.01) ^{Ab}	9.78 (0.01) ^{Ab}	7.65 (0.02) ^{Ac}	7.65 (0.02) ^{Ac} 7.83 (0.04) ^{Bb} 8.65 (0.02) ^{Ba}	$8.65 (0.02)^{Ba}$
10% aTCP	315	7.72 (0.02) ^{Ab}	7.72 (0.02) ^{Ab} 7.78 (0.13) ^{Bb} 8.63 (0.10) ^{Ba}	$8.63 (0.10)^{Ba}$	7.64 (0.03) ^{Ac}	7.64 (0.03) ^{Ac} 7.74 (0.00) ^{Cb} 8.37 (0.01) ^{Ca}	8.37 (0.01) ^{Ca}
15% aTCP	405	7.59 (0.01) ^{Aa}	7.59 (0.01) ^{Aa} 7.53 (0.10) ^{Ca} 7.55 (0.15) ^{Ca}	$7.55 (0.15)^{Ca}$	7.55 (0.01) ^{Bc}	7.55 (0.01) ^{Bc} 7.77 (0.02) ^{Cb} 8.21 (0.04) ^{Da}	8.21 (0.04) ^{Da}
Initial pH of w	ater and SBF is	itial pH of water and SBF is 5.78 (0.12) and 7.40 (0.01), respectively. Means followed by distinct capital letters indicates	.40 (0.01). respe	ectively. Means	followed by dist	inct capital let	ters indicates

significant difference (p<0.05) for the same storage period. Means followed by distinct lowercase letters indicates significant

difference (p<0.05) between pH measured in different times

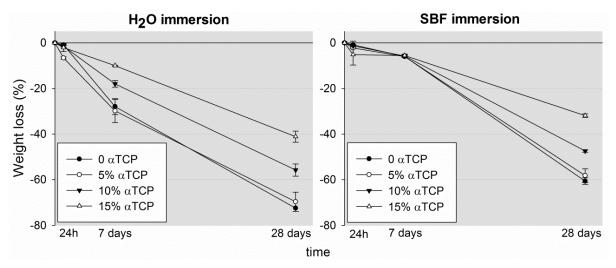


Figure 1. Weight loss (%) of the sealers after soaking in water or SBF for different periods.

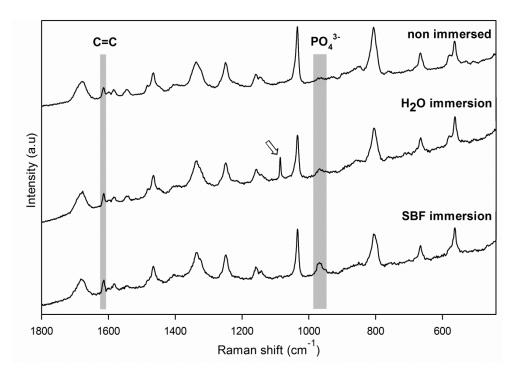


Figure 2. Representative Raman spectra of 15% α TCP sealer after setting and after 7 days of water or SBF immersion. The 1613- and 962-cm⁻¹ regions in vibration mode, carbon-to-carbon stretching of the aromatic ring of salicylic group, and a symmetric stretching mode of phosphate, respectively, were highlighted. Note the presence of a peak at 1084 cm⁻¹ (open arrow) on the spectra of a sealer after water immersion. This peak corresponds to the Ca(OH)₂ chemical bonds and could repeatedly be viewed for all groups after soaking specimens in water.

A representative Raman spectrum is shown in Figure 2. The peak corresponding to the stretching mode of phosphate increases after seven days of immersion in distillated water and SBF. A peak at 1084 cm⁻¹ was noticed for specimens immersed in water. Figure 3 shows a mapping characterization of the specimen's surface regarding the changes in ratio of the intensity of 962-cm⁻¹ and 1613-cm⁻¹ Raman bands ($A_{962cm}^{-1}/A_{1613cm}^{-1}$). In the non-immersed specimens, the ratio $A_{962cm}^{-1}/A_{1613cm}^{-1}$ increased with the increased amount of α TCP in the sealer's composition. After water and SBF immersion, no changes were seen in the 0%and 5% α TCP groups, while for 10% and 15% α TCP compositions, changes occurred, with an increase in the $A_{962cm}^{-1}/A_{1613cm}^{-1}$ ratio.

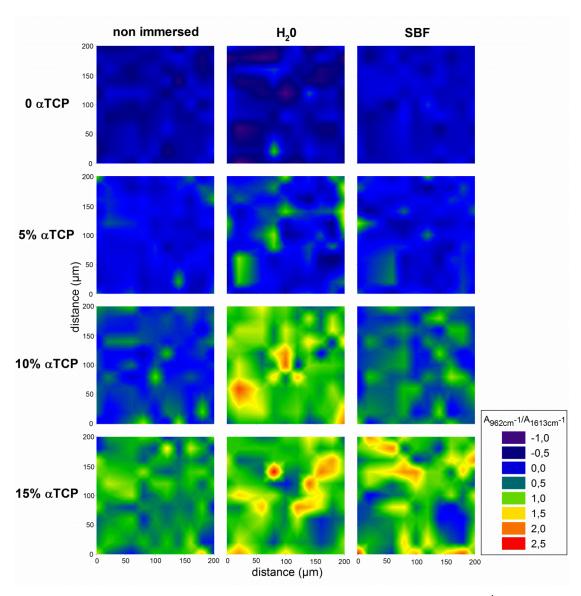


Figure 3. Raman mapping of the intensity of the ratio of the 962-cm⁻¹ peak area to the 1613-cm⁻¹ peak area measured on a 200x200-µm area of sample surfaces.

The values of the $A_{962cm}^{-1}/A_{1613cm}^{-1}$ ratios are presented in Figure 4. In the 10% α TCP sealer, the initial value of 0.32 increased to 0.88 for specimen immersion in water and 0.45 for SBF. The 15% α TCP sealer ratio ranged from 0.66 to 0.90 and 0.87, after water and SBF immersion, respectively.

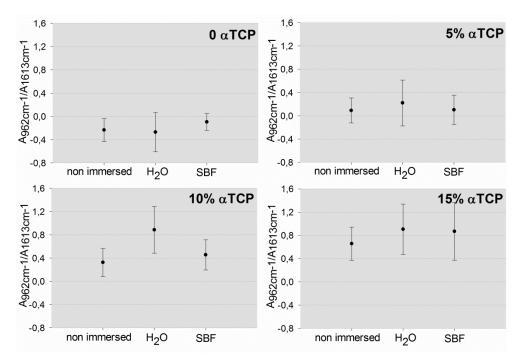


Figure 4. Variation of the ratio of the 962-cm⁻¹ peak area to the 1613-cm⁻¹ peak area measured on Raman spectra in function of specimen treatment (non-immersed, after water storage and after SBF storage) for each sealer. The circle represents the mean value of 100 points, and the bars represent the standard deviation.

Figure 5 presented the microstructure of sealers after 7 days of immersion in distillated water and SBF. SEM images revealed that all sealers present similar surface morphology before immersion. The EDX analysis after water immersion showed an increase in calcium and phosphorus on the specimen surface. The immersion in SBF produced irregular surfaces with calcium and phosphorous at elevated concentrations, except for the 15% α TCP. A representative image of a needle-like structure found around calcium and phosphorous deposition is shown in Figure 6. Figure 7 shows fibroblast viability after 24 or 48 h in direct contact with sealers. The sealers with α TCP additionally presented lower cell viability than commercial sealer AHPlus at 24 h (p<0.05). However, the incorporation of α TCP showed no difference from the commercial sealer (p>0.05).

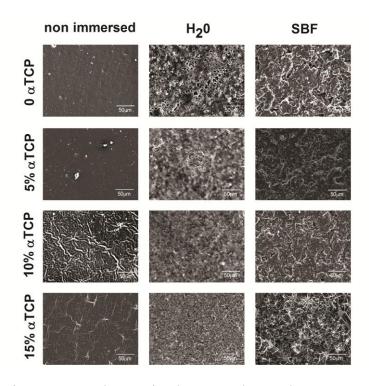


Figure 5. SEM (500x magnification) of the surface of each sealer after different treatments.

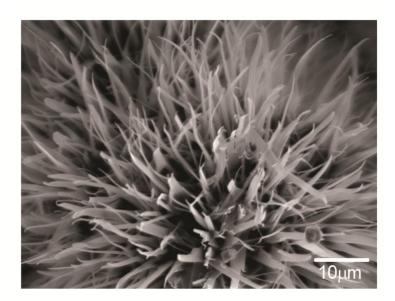


Figure 6. SEM (2000x magnification) of 15% α TCP sealer after SBF immersion for 7 days.

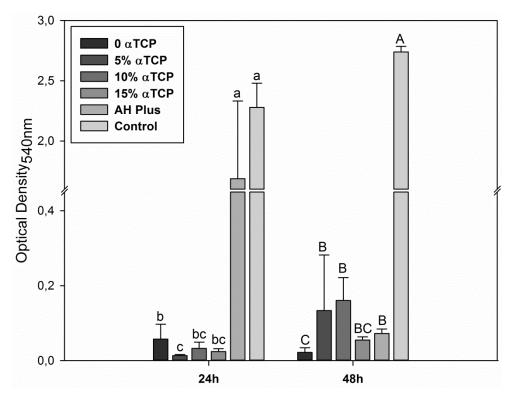


Figure 7. Cell viability (±standard deviation) of L929 upon direct contact with sealers after culturing for 24 h and 48 h. Distinct lowercase letters indicate a significant difference (p<0.05) at 24 h, and distinct capital letters indicate a difference at 48 h.

DISCUSSION

Grossman's concept¹⁹ posits that inert materials are preferred to root canal sealers (discussed recently). New bioactive root canal sealers introduced in clinical use accelerate healing of apical tissues and promote effective sealing of the root canal system^{20, 21}. The α -tricalcium phosphate sealers engineered in this study presented bioactive potential to be used in periapical teeth tissues. The addition of α TCP increased the experimental glycerol salicylate root canal sealer properties.

Hardening of the salicylate sealers is promoted by calcium chelation achieved by a 1:5 ratio of calcium hydroxide to $resin^7$. To ensure correct stoichiometry of the chemical reaction, 30%wt of inorganic compounds with at least 15% calcium hydroxide was used for the development of sealers. The setting reaction time ranged from 240 to 405 min, similar to commercial sealers²². The faster setting time was recorded at 0% and 5% α TCP groups (30% and 25% calcium hydroxide, respectively). The high reactivity and availability of calcium hydroxide in comparison to α TCP may explain the faster reaction obtained. Further, the conversion of α TCP

into a calcium-deficient hydroxyapatite (CDHA) by hydration^{15, 23} could retard the sealer setting, as in the following equation:

$$3 \alpha Ca_3(PO_4)_2 + H_2O \rightarrow Ca_9(HPO_4)(PO_4)_5OH.$$

Groups with 0% and 5% α TCP presented the same setting time and increased pH, probably due to a fraction of calcium hydroxide not reacting that was quickly released. The pH increased for all groups after 28 days. As the concentration of α TCP decreased, an increase in pH was observed due to decreased hydroxyl ion release. The optimum pH of alkaline phosphatase activity in bone formation is 8.5^{24} . In this study, the pH achieved by α TCP sealers was approximately 60% of enzymatic activity²⁵. The alkalization potential shown by the experimental sealers could lead to an increased rate of healing process, antimicrobial properties and biocompatibility. The cytotoxicity of experimental sealers was higher than that of the commercial sealer in 24 h, reaching no significant difference after 48 h. These results could be due to an increased setting after 48 h and to a regeneration process of fibroblast cells. The cell viability values measured are in accordance with other studies evaluating several root canal sealers^{26, 27} via direct contact with fibroblast cells.

Root canal sealer specimens immersed in SBF presented less weight loss than in water. The SBF ion-saturated medium could have promoted the conversion of α TCP in CDHA, decreasing the calcium phosphate compound solubility²⁸, which could be observed by the decreased weight loss with increased α TCP concentration. After 28 days in SBF, the 15% α TCP sealers exhibited 31.86% loss in mass in contrast with 60.63% of the 0 α TCP sealer in the same period. Faria-Júnior *et al.*⁶ evaluated the solubility of salicylate-based sealers after 15 h of water immersion with values ranging from 9.97% to 15.03%. The authors showed that higher solubility leads to increased pH and efficacy against *Enterococcus faecalis* biofilm.

A requirement for biomaterials is the capacity to promote the deposition of bone-like apatite on the surface when implanted in a living body²⁹. Interaction between the bioactive sealer and fluid tissues has been viewed as a promising approach to heal a compromised seal²¹. In this study, sealers with 10% and 15% α TCP showed a noticeable phosphate content on the surface independent of immersion media. Water presented in a medium leads to the transformation of α TCP into CDHA¹⁵, and a release of Ca²⁺ and PO₄³⁻ could lead to nucleation and crystallization as apatite. In water-immersed specimens, an erosion process could be observed (Fig. 5)³⁰ as the calcium hydroxide concentration increased. Groups with

15% αTCP showed no surface erosion due to a decreased content of calcium hydroxide. The peak of calcium hydroxide (1084cm⁻¹) was present in the Raman spectra of water immersed specimens⁷. After SBF immersion, specimens presented an irregular surface with grains with higher calcium and phosphate concentrations than the non-immersed samples, indicating an apatite layer growth. Szuber et al. 16 noted similar change patterns for α-tricalcium phosphate cement after SBF immersion. The SEM-EDX showed the early formation of a calcium-phosphate layer on a sealer's surface. The calcium phosphate formed on a sample surface suggests the bioactivity of aTCP sealers. In vivo, the clastic cells of periapical tissue could partially dissolve this calcium-phosphate layer, releasing Ca²⁺ and PO₄³⁻, leading to the precipitation of carbonated hydroxyapatite due to supersaturation. The precipitation of stains and crystallites of calcium phosphate at voids between core materials and sealer could enhance the sealing. A study showed that calcium silicate-based sealers are capable of promoting the precipitation of calciumphosphate deposits on interfacial voids of obturation materials, reducing the empty spaces and improving the sealing ability in comparison to traditionally used sealers³¹. Furthermore, the apatite layer formed could facilitate protein adsorption and osteprogenitor cell attachment. The calcium and phosphate ions released trigger the osteoblastic proliferation and differentiation to achieve biomineralization 32, 33, leading to a biological seal.

Therefore, the experimental root canal sealer obtained from the mixture of glycerol salicylate resin, calcium hydroxide and α -tricalcium phosphate showed solubility, hydroxyl ion release and surface changes in the formation of an apatite layer suitable to be used as a bioactive sealer for root canal filling. Thus, glycerol salicylate sealers with 10% and 15% α -tricalcium phosphate showed reliable physical chemical properties and apatite-forming ability.

ACKNOWLEDGMENTS

FFP gratefully acknowledges scholarship support from CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior). The authors thank CME-UFRGS for support for SEM images acquisition.

REFERENCES

- Ng YL, Mann V, Gulabivala K. A prospective study of the factors affecting outcomes of nonsurgical root canal treatment: part 1: periapical health. Int Endod J 2011;44:583-609.
- 2. Tyagi S, Mishra P, Tyagi P. Evolution of root canal sealers: An insight story. Eur J Gen Dent 2013;2:199-218.
- 3. Waltimo TM, Boiesen J, Eriksen HM, Orstavik D. Clinical performance of 3 endodontic sealers. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2001;92:89-92.
- 4. Amann R, Peskar BA. Anti-inflammatory effects of aspirin and sodium salicylate. Eur J Pharmacol 2002;447:1-9.
- Leonardo MR, Silva LA, Utrilla LS, Assed S, Ether SS. Calcium hydroxide root canal sealers-histopathologic evaluation of apical and periapical repair after endodontic treatment. J Endod 1997;23:428-32.
- Faria-Junior NB, Tanomaru-Filho M, Berbert FL, Guerreiro-Tanomaru JM. Antibiofilm activity, pH and solubility of endodontic sealers. Int Endod J 2013;46:755-62.
- Portella FF, Santos PD, Lima GB, Leitune VC, Petzhold CL, Collares FM, Samuel SM. Synthesis and characterization of a glycerol salicylate resin for bioactive root canal sealers. Int Endod J 2014; 47:339-45.
- 8. Bae WJ, Chang SW, Lee SI, Kum KY, Bae KS, Kim EC. Human periodontal ligament cell response to a newly developed calcium phosphate-based root canal sealer. J Endod 2010;36:1658-63.
- Bae KH, Chang SW, Bae KS, Park DS. Evaluation of pH and calcium ion release in capseal I and II and in two other root canal sealers. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2011;112:e23-8.
- Shokouhinejad N, Nekoofar MH, Razmi H, Sajadi S, Davies TE, Saghiri MA, Gorjestani H, Dummer PM. Bioactivity of EndoSequence root repair material and bioaggregate. Int Endod J 2012;45:1127-34.
- 11. Yang SE, Baek SH, Lee W, Kum KY, Bae KS. In vitro evaluation of the sealing ability of newly developed calcium phosphate-based root canal sealer. J Endod 2007;33:978-81.

- 12. dos Santos LA, Carrodeguas RG, Rogero SO, Higa OZ, Boschi AO, de Arruda AC. Alpha-tricalcium phosphate cement: "in vitro" cytotoxicity. Biomaterials 2002;23:2035-42.
- 13. ISO 6876 Dental root canal sealing materials. Geneva, Switzerland: International Organization for Standardization; 2001.
- 14. Kokubo T, Takadama H. How useful is SBF in predicting in vivo bone bioactivity? Biomaterials 2006;27:2907-15.
- 15. Morejon-Alonso L, Ferreira OJ, Carrodeguas RG, dos Santos LA. Bioactive composite bone cement based on alpha-tricalcium phosphate/tricalcium silicate. J Biomed Mater Res B Appl Biomater 2012;100:94-102.
- 16. Szubert M, Adamska K, Szybowicz M, Jesionowski T, Buchwald T, Voelkel A. The increase of apatite layer formation by the poly(3-hydroxybutyrate) surface modification of hydroxyapatite and beta-tricalcium phosphate. Mater Sci Eng C Mater Biol Appl 2014;34:236-44.
- 17. Dain RP, Gresham G, Groenewold GS, Steill JD, Oomens J, van Stipdonk MJ. Infrared multiple-photon dissociation spectroscopy of group II metal complexes with salicylate. Rapid Commun Mass Spectrom 2011;25:1837-46.
- ISO 10993-5 Biological evaluation of medical devices: Tests for in vitro cytotoxicity. Geneva, Switzerland: International Organization for Standardization; 2009.
- 19. Gatewood RS. Endodontic materials. Dent Clin North Am 2007;51:695-712.
- 20. Alani A, Knowles JC, Chrzanowski W, Ng YL, Gulabivala K. Ion release characteristics, precipitate formation and sealing ability of a phosphate glasspolycaprolactone-based composite for use as a root canal obturation material. Dent Mater 2009;25:400-10.
- 21. Li GH, Niu LN, Zhang W, Olsen M, De-Deus G, Eid AA, Chen JH, Pashley DH, Tay FR. Ability of new obturation materials to improve the seal of the root canal system: A review. Acta Biomater 2014;10:1050-63.
- 22. Zhou HM, Shen Y, Zheng W, Li L, Zheng YF, Haapasalo M. Physical properties of 5 root canal sealers. J Endod 2013;39:1281-6.
- 23. Ginebra MP, Fernandez E, De Maeyer EA, Verbeeck RM, Boltong MG, Ginebra J, Driessens FC, Planell JA. Setting reaction and hardening of an apatitic calcium phosphate cement. J Dent Res 1997;76:905-12.

- 24. Orimo H. The mechanism of mineralization and the role of alkaline phosphatase in health and disease. J Nippon Med Sch 2010;77:4-12.
- 25. Harada M, Udagawa N, Fukasawa K, Hiraoka BY, Mogi M. Inorganic pyrophosphatase activity of purified bovine pulp alkaline phosphatase at physiological pH. J Dent Res 1986;65:125-7.
- 26. Brackett MG, Messer RL, Lockwood PE, Bryan TE, Lewis JB, Bouillaguet S, Wataha JC. Cytotoxic response of three cell lines exposed in vitro to dental endodontic sealers. J Biomed Mater Res B Appl Biomater 2010;95:380-6.
- 27. Brackett MG, Marshall A, Lockwood PE, Lewis JB, Messer RL, Bouillaguet S, Wataha JC. Cytotoxicity of endodontic materials over 6-weeks ex vivo. Int Endod J 2008;41(12):1072-8.
- 28. Dorozhkin SV. Calcium orthophosphates in dentistry. J Mater Sci Mater Med 2013;24:1335-63.
- 29. Kokubo T. Bioactive glass ceramics: properties and applications. Biomaterials 1991;12:155-63.
- 30. Akashi A, Matsuya Y, Nagasawa H. Erosion process of calcium hydroxide cements in water. Biomaterials 1991;12:795-800.
- 31. Gandolfi MG, Parrilli AP, Fini M, Prati C, Dummer PM. 3D micro-CT analysis of the interface voids associated with Thermafil root fillings used with AH Plus or a flowable MTA sealer. Int Endod J 2013;46:253-63.
- 32. LeGeros RZ. Calcium phosphate-based osteoinductive materials. Chem Rev 2008;108:4742-53.
- 33. Samavedi S, Whittington AR, Goldstein AS. Calcium phosphate ceramics in bone tissue engineering: a review of properties and their influence on cell behavior. Acta Biomater 2013;9:8037-45.

4. CONSIDERAÇÕES FINAIS

Esforços visando ao desenvolvimento de novos cimentos obturadores que propiciem um melhor vedamento em comparação aos materiais disponíveis no mercado, são necessários, uma vez que o selamento do sistema de canais radiculares tem papel fundamental no sucesso da terapia endodôntica. Nesse estudo foi proposto um cimento obturador à base de uma resina de glicerol salicilato e α-fosfato tricálcico. Espera-se que o emprego da resina de glicerol salicilato produza um material com menor solubilidade, quando comparado aos cimentos comerciais contendo resina de isobutil salicilato, e que a adição de α-fosfato tricálcico confira bioatividade ao cimento, de forma que a interação com os tecidos periapicais leve a uma deposição mineral da superfície do material, melhorando o selamento e estimulando o reparo tecidual.

Os objetivos propostos na presente tese foram alcançados e estão contemplados em dois artigos, de forma que o Artigo I mostra o processo de obtenção da nova resina de salicilato e confirma a sua aplicabilidade para uso em cimentos que contenham íons cálcio disponíveis para reagir com os grupos salicilato da resina, auferindo presa química. No Artigo II foram formulados cimentos a partir da resina sintetizada e da adição de hidróxido de cálcio e α-fosfato tricálcico, em diferentes concentrações. Observou-se que maiores concentrações de α-fosfato tricálcico (10% e 15%) produziram cimentos mais resistentes à degradação em água e com potencial de estimular a deposição de fosfatos de cálcio sobre a sua superfície.

Espera-se que a precipitação mineral, observada *in vitro* na superfície dos discos obtidos com o cimento desenvolvido, quando empregado para obturação de canais radiculares também ocorra na interface obturação/dentina e na região periapical, otimizando o selamento apical, e consequentemente a taxa de sucesso da terapia. Os resultados favoráveis obtidos nestes trabalhos justificam plenamente mais investimentos na avaliação e ajuste, se necessário, das propriedades de escoamento, espessura de película e radiopacidade, de acordo com a ISO 6876 (Dental root canal sealing materials), para viabilizar o uso clínico desse material. Além disso, as propriedades físicas e químicas do cimento podem ser ajustadas

para que o mesmo possa ser, utilizado também para proteção pulpar, haja vista que a alcalinização e a deposição mineral promovidas pelo material são desejadas tanto para a formação de dentina reacional quanto para a remineralização dentinária, nos casos de remoção parcial de tecido cariado.

REFERÊNCIAS

ALANI, A., *et al.* Ion release characteristics, precipitate formation and sealing ability of a phosphate glass-polycaprolactone-based composite for use as a root canal obturation material. **Dent Mater**, v.25, n.3, p.400-10. 2009.

BERBERT, F. L., *et al.* Influence of root canal dressings and sealers on repair of apical periodontitis after endodontic treatment. **Oral Surg Oral Med Oral Pathol Oral Radiol Endod**, v.93, n.2, p.184-9. 2002.

CHI, Z. Y., *et al.* A laboratory study of producing docosahexaenoic acid from biodiesel-waste glycerol by microalgal fermentation. **Process Biochemistry**, v.42, n.11, p.1537-45. 2007.

DOROZHKIN, S. V. Bioceramics of calcium orthophosphates. **Biomaterials**, v.31, n.7, p.1465-85. 2010.

DOROZHKIN, S. V. Calcium orthophosphates in dentistry. **J Mater Sci: Mater Med**, v.24, n.6, p.1335-63. 2013.

FARIA-JUNIOR, N. B., *et al.* Antibiofilm activity, pH and solubility of endodontic sealers. **Int Endod J**, v.46, n.8, p.755-62. 2013.

FLUHR, J. W., *et al.* Glycerol and the skin: holistic approach to its origin and functions. **Br J Dermatol**, v. 159, n.1, p.23-34. 2008.

GANDOLFI, M. G., *et al.* Apatite formation on bioactive calcium-silicate cements for dentistry affects surface topography and human marrow stromal cells proliferation. **Dent Mater**, v. 26, n.10, p. 974-92. 2010.

GILLEN, B. M., *et al.* Impact of the quality of coronal restoration versus the quality of root canal fillings on success of root canal treatment: a systematic review and meta-analysis. **J Endod**, v.37, n.7, p.895-902. 2011.

GINEBRA, M. P., *et al.* Setting reaction and hardening of an apatitic calcium phosphate cement. **J Dent Res**, v.74, n.4, p. 905-12. 1997.

GUASTALDI, A. C.; APARECIDA, A. H. Fosfatos de cálcio de interesse biológico: importância como biomateriais, propriedades e métodos de obtenção de recobrimentos. **Quim. Nova**, v.33, n.6, p.1352-8. 2010.

HERNANDEZ-IZQUIERDO, V. M.; Krochta, J. M. Thermoplastic processing of proteins for film formation-a review. **J Food Sci**, v.73, n.2, p.R30-9. 2008.

ISO 6876. Dental root canal sealing materials. Genebra, 2012.

KARINEN, R. S.; KRAUSE, A. O. I. New biocomponents from glycerol. **Applied Catalysis a-General**, v.306, p.128-33. 2006.

KIM, J., *et al.* Functional biomimetic analogs help remineralize apatite-depleted demineralized resin-infiltrated dentin via a bottom-up approach. **Acta Biomater**, v.6, n.7, p.2740-50. 2010.

LEONARDO, M., *et al.* Calcium hydroxide root canal sealers: histopathologic evaluation of apical and periapical repair after endodontic treatment. **J Endod**, v.23, n.7, p.428-32. 1997.

LI B., *et al.* Effect of nanostructure on osteoinduction of porous biphasic calcium phosphate ceramics. **Acta Biomater**, v.8, n.10, p.3794-804. 2012.

LIU, Y., *et al.* Differences between top-down and bottom-up approaches in mineralizing thick, partially demineralized collagen scaffolds. **Acta Biomater**, v.7, n.4, p.1742-51. 2011.

MAI, S., *et al.* In vitro remineralization of severely compromised bonded dentin. **J Dent Res**, v.89, n.4, p.405-10. 2010.

MIGNECO, F.; HUANG, Y. C.; BIRLA, R. K.; HOLLISTER, S. J. Poly(glycerol-dodecanoate), a biodegradable polyester for medical devices and tissue engineering scaffolds. **Biomaterials**, v.30, n.33, p.6479-84. 2009.

MOREJON-ALONSO, L., *et al.* Bioactive composite bone cement based on alphatricalcium phosphate/tricalcium silicate. **J Biomed Mater Res B Appl Biomater**, v.100, n.1, p.94-102. 2012.

Ng, Y. L.; Mann, V.; Gulabivala, K. A prospective study of the factors affecting outcomes of nonsurgical root canal treatment: part 1: periapical health. **Int Endod J**, 2011;44:583-609.

PORTELLA, F. F., *et al.* Synthesis and characterization of a glycerol salicylate resin for bioactive root canal sealers. **Int Endod J**, v.47, n.4, p.339-45. 2014.

PORTELLA, F. F., *et al.* Glycerol salicylate-based containing α-tricalcium phosphate as a bioactive root canal sealer. **J Biomed Mater Res B Appl Biomater**, 2015.

SALVI, B. L.; PANWAR, N. L. Biodiesel resources and production technologies – A review. **Renewable and Sustainable Energy Reviews**, v.16, p.3680-9. 2012.

SILVA, L. A., *et al.* Inflammatory response to calcium hydroxide based root canal sealers. **J Endod**, v.23, n.2, p.86-90. 1997.

The Kerr Corporation. Waller D.E.; Jandourek E. **Dental Resins**. USA n. 5922785, 13 jul. 1999.

WALTIMO, T. M., *et al.* Clinical performance of 3 endodontic sealers. **Oral Surg Oral Med Oral Pathol Oral Radiol Endod**, v.92, n.1, p.89-92. 2001.

YUCEL, A. C., *et al.* Bacterial penetration after obturation with four different root canal sealers. **J Endod**, v.32, n.9, p.890-3. 2006.

CIP - Catalogação na Publicação

Freitas Portella, Fernando

Cimento experimental bioativo à base de glicerol salicilato contendo alfa-fosfato tricálcico para uso endodôntico / Fernando Freitas Portella. -- 2015. 51 f.

Orientadora: Susana Maria Werner Samuel.

Tese (Doutorado) -- Universidade Federal do Rio Grande do Sul, Faculdade de Odontologia, Programa de Pós-Graduação em Odontologia, Porto Alegre, BR-RS, 2015.

1. Cimentos dentários. 2. Fosfatos de cálcio. 3. Hidróxido de cálcio. 4. Obturação do canal radicular. I. Werner Samuel, Susana Maria, orient. II. Título.