### UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL FACULDADE DE FARMÁCIA TRABALHO DE CONCLUSÃO DE CURSO EM FARMÁCIA

Potential additive or synergistic effect of the essential oils of *Eucalyptus citriodora*, *Eucalyptus camaldulensis* and *Eucalyptus globulus* and their interactions with antifungal agents to evaluate anti-*Candida* spp. activity: a literature review

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# Potential additive or synergistic effect of the essential oils of *Eucalyptus citriodora*, *Eucalyptus camaldulensis* and *Eucalyptus globulus* and their interactions with antifungal agents to evaluate anti-*Candida* spp. activity: a literature review

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#### **Abstract**

The genus *Candida* covers a diversity of species responsible for causing important fungal infections in individuals. *Candida* species are among the most frequent pathogens in hospital infections considered severe. The increasing resistance to antifungal drugs is one of the factors that promote prospecting for new therapeutic agents. Essential oils have shown promising results by inhibiting or preventing fungal growth.

A literature review was performed in the online databases PubMed, Scielo, Scopus, LILACS, CAPES periodicals and ScienceDirect, with the aim of verifying the anti-candid activity and possible interactions with antifungals of essential oils of *Eucalyptus citriodora*, *Eucalyptus camaldulensis* and *Eucalyptus globulus*. The main constituents of these essential oils are citronellal, 1,8-cineole (eucalyptol), have anti-*Candida* spp. activity and have a potential additive or synergistic effect when combined with antifungals. The MIC range of *E. citriodora* essential oil for different *Candida* species was 0,02 μg/mL to 5 μg/mL. The range of the inhibition zone of the essential oil of *E. camaldulensis* against the different species of *Candida* was 18 – 23 mm for the leaves of the plant and 12 - 20 mm for the fruits. The MIC for the essential oil of E. globulus was 1000 μg/mL, while in combination with an antifungal, the value was 32 times lower, thus presenting an additive effect with ICIF of 1.031. They also have a potential additive or synergistic effect with antimicrobials. However, further studies are still needed to consolidate knowledge about these species for their use in the therapeutic clinic in infections caused by *Candida* spp.

Keywords: Candida; Eucalyptus; antifungals; essential oils; synergism; additive effect.

#### Introduction

Candida is a yeast present in the normal microbiota of humans. It is present in the oral cavity, genital tract, perianal region and gastrointestinal tract. The presence of Candida does not necessarily imply infection. However, when there is an imbalance in defense mechanisms or anatomical barriers, especially in individuals with immunosuppression, it may become pathogenic. Some factors may contribute to the development of the disease, such as malnutrition, obesity, diabetes, pregnancy, antibiotic therapy, chemotherapy, corticosteroid use, neoplasms and other debilitating diseases [1, 2].

Candidiasis is an opportunistic mycosis caused by *Candida* spp. with varied clinical manifestations, which may cause superficial, cutaneous, subcutaneous and systemic infections. Acute and chronic infections manifest with lesions in the mouth, pharynx, skin, nails, bronchopulmonary system, intestinal, perianal, and occasionally, endocarditis, meningitis, fungemia or infections elsewhere [1, 2].

Treatment of candidiasis is done according to the clinical manifestations of the patient and the severity of the disease. In superficial candidiasis, imidazole derivatives such as clotrimazole, miconazole, ketoconazole, oxyconazole, terconazole and polyenics such as nystatin are used. In more extensive cases, systemic drugs such as itraconazole or fluconazole are administered. In systemic candidiasis, amphotericin B is also one of the drugs chosen, usually in association with 5 fluorocytosine or other drugs such as triazole derivatives, fluconazole and itraconazole [3, 4].

According to the Center for Diseases Control and Prevention (CDC, 2020) some *Candida* species have demonstrated resistance to commonly used antifungal drugs. About 7% of all *Candida* spp. infected blood samples tested at CDC are fluconazole resistant. In addition, the resistance to the class of echinocandins by *Candida glabrata* has become worrisome due

to the fact that it also presents high rates of resistance to fluconazole. This limits the therapeutic options for patients infected by this species. Another species that has been resistant to antifungals is *Candida auris*. About 90% of *C. auris* samples in the USA were resistant to fluconazole and 30% resistant to amphotericin B [5]. Other studies report intrinsic resistance to antifungals in species of *Candida*. *C. krusei*, intrinsically resistant to fluconazole and some other *Candida* species have intrinsic resistance to amphotericin B [6, 7].

Thus, prospecting for new therapeutic agents for the treatment of candidiasis is increasingly important. Natural products have been used in practice for many years to control and cure diseases. Currently, plant essential oils have been studied by several researchers around the world and have shown promise. Several studies have shown that some essential oils may present antifungal activity in their constituents, isolated or not, and may also have a synergistic effect when in use with antimicrobial drugs. A synergistic effect between the two compounds can reduce the dose to be used and thus reduce the toxicity and possible side effects of both compounds, representing an alternative to the resistance of microorganisms [8, 9].

Several essential oils are reported in the literature with antifungal activity. The species of the genus *Eucalyptus* have a smaller number of studies when compared to other plant genera. In this sense, this review aims to verify the studies conducted with the essential oils of *Eucalyptus citriodora*, *Eucalyptus camaldulensis* and *Eucalyptus globulus* for chemical composition, anti-*Candid*a spp. activity and possible pharmacological interactions with antifungals used in therapeutic clinic.

#### Methodology

This study constitutes a review of the anti-Candida spp. activity by the essential oils of Eucalyptus citriodora, Eucalyptus camaldulensis and Eucalyptus globulus and the possible interactions with antifungals. The databases of the Scientific Electronic Online Library (SciELO), National Library of Medicine (PubMed), Scopus, LILACS, CAPES periodicals and Science Direct were used for the research. The searches were carried out from May to August 2020. The inclusion criteria were scientific articles addressing the proposed theme published between 2015 and 2020.

The first search was conducted with the combinations of words "antifungal AND oils AND Candida AND synergism" to list the essential oils being studied in the last five years. After that, a second survey was conducted to search for data on Eucalyptus species, using the combinations of words "antifungal AND oils AND Candida AND synergism AND Eucalyptus camaldulensis; antifungal AND oils AND Candida AND synergism AND Eucalyptus citriodora; antifungal AND oils AND Candida AND synergism AND Eucalyptus globulus".

The chemical composition of *E. citriodora*, *E. camaldulensis* and *E. globulus* was researched under the same conditions, using the combinations of words "*Eucalyptus citriodora* AND composition; *Eucalyptus camaldulensis* AND composition; *Eucalyptus globulus* AND composition; *Eucalyptus citriodora* AND oil essential; *Eucalyptus camaldulensis* AND oil essential; *Eucalyptus globulus* oil essential". In this search, there were no criteria of choice for the selection of articles in relation to the regions of cultivation, seasons or extraction methods of the species of interest.

The anti-*Candida* spp. activity of the two species of *E. citriodora*, *E. camaldulensis* and *E. globulus* was researched in the databases maintaining the five-year filter, using the combinations of words "*Eucalyptus citriodora* AND *Candida*; *Eucalyptus camaldulensis* AND *Candida*; *Eucalyptus globulus* AND *Candida*".

The evaluation of the interactions of essential oils and their main constituents of both species of *E. citriodora*, *E. camaldulensis* and *E. globulus* with antifungals against *Candida* spp. was performed using the combinations of words "checkerboard AND *Eucalyptus* 

citriodora; checkerboard AND Eucalyptus camaldulensis; checkerboard AND Eucalyptus globulus; checkerboard AND citronellal; checkerboard AND 1,8 cineole; checkerboard AND eucalyptol".

#### **Discussion**

Several plants are used in the therapeutic routine for the treatment of different diseases. The essential oils of some plants, alone or in combination, are being studied to prove their activities against different pathogens. The genus *Candida* spp. includes species responsible for causing severe infections and, in addition, some species have been presented to resistance to commonly used antifungals. Thus, the present study evaluates the anti-*Candida* spp. activity by the essential oils of *Eucalyptus citriodora*, *Eucalyptus camaldulensis* and *Eucalyptus globulus*, as well as the possible interactions with antifungals, in order to contribute to the therapeutic arsenal the infections caused by *Candida* spp. The first search generated a total of 496 newspapers.

Duplicate and tripiclata newspaper were excluded. The others were selected for title and abstract reading to verify the relevance of the study. After reading the titles and abstracts it was possible to list the species that are being studied in the last 5 years. Each species listed was searched in the databases to analyze, in numbers, the relevance of each species. *Thymus vulgaris* appears at the top of the list, being cited in 143 articles. The *Origanum vulgare* is the second most researched, being cited in 108 articles. *Citrus limonum* was cited in 86 articles.

The plant species most used for the evaluation of anti-Candida spp. activity are the essential oils of Thymus vulgaris, Origanum vulgare and Citrus limonum. The species Eucalyptus camaldulensis was cited in 26 articles and Eucalyptus citriodora was mentioned in 18 articles.

#### 1. Essential oils with anti-Candida spp. activity

In recent years, the antifungal activity of several plant species has been studied. The most researched essential oils against *Candida* spp. are thyme *(Thymus vulgaris)*, oregano *(Origanum vulgare)*, lemon *(Citrus limonum)*, clove *(Syzygium aromaticum)*, melaleuca *(Melaleuca alternifolia)*, cinnamon *(Cinnamomum verum)*, mint *(Mentha piperita)* and others.

Gukwa K et al. [10] reported fungistatic and fungicide activity in relation to C. albicans and C. glabrata for the essential oils of Thymus vulgaris, Citrus limonum, Pelargonium graveolens, Cinnamomum cassia, Ocimum basilicum and Eugenia caryophyllus. The best activity was observed for essential oil of Cinnamomum cassia.

Mandras N *et al.* [11] demonstrated that the essential oils *of Thymus vulgaris* (thyme), *Foeniculum vulgare* (fennel), *Eugenia caryophyllata* (carnation), *Pinus sylvestris* (pine), *Salvia officinalis* (sage), Melissa *officinalis* (lemon grass) and *Lavandula vera* (lavender) have good activity against *Candida* sp. The higher activity of thyme and pine essential oils may be related to its main components, carvacrol, thymol and  $\alpha$ -pinene, which have proven antifungal action.

Cardoso NNR *et al.* [12] reported the antifungal activity of the essential oil of *Ocimum basilicum* and its main components against *Candida albicans*. The best results were presented by the geraniol component.

Bhat V et al. [13] proved that *Origanum vulgare* essential oil has high anti-*Candida* spp. properties against oral clinical isolates. *O. vulgare* showed much lesser MIC/MFC as compared to fluconazole indicating that the herb can be effective even in a lower dose of MIC/MFC. *O. vulgare* showed much lower MIC/MFC values compared with fluconazole indicating that essential oil may be effective even at a lower dose of MIC/MFC. The main component of *O. vulgare* is the carvacrol, which has proven antifungal activity.

#### 2. Pharmacological interactions of antifungals against Candida spp.

Recent studies have shown that essential oils extracted from plants can have a synergistic effect when associated with an antimicrobial agent. Different authors report synergism between essential oil constituents and antifungal drugs. De Castro RD *et al.* [14] showed that the thymol has a fungicide effect on the species *Candida* and also has a synergistic effect with nystatin. Gukwa K *et al.* [10] observed that the essential oils of *Pelargonium graveolens* and *Cinnamomum cassia* showed synergistic activity with amphotericin B against *C. albicans* and *C. glabrata*. Cardoso NNR *et al.* [12] reported synergistic activity of geraniol and linalool, essential oil components of *Ocimum basilicum*, with fluconazole, especially against fluconazole-resistant *C. albicans* strain.

#### 3. Eucalyptus citriodora, Eucalyptus camaldulensis and Eucalyptus globulus

The genus *Eucalyptus* contains more than 800 species and is one of the most used plants in the world. Different parts of the plant are used in various areas such as in the pulp and paper industry, food industry, dentistry and medicine. More than 300 species of this genus contain volatile oils in their leaves, being used as fragrance elements in household and cosmetic products such as soaps, detergents, lotions and perfumes. They are also used as flavor elements in food and beverages. In addition, in clinical practice *Eucalyptus* is used to prevent and treat human diseases due to its antimicrobial, anti-inflammatory and antioxidant properties. *Eucalyptus* essential oil is present in different formulations, such as syrups, tablets and ointments. It is widely used as expectant for cough, in burns, as muscle relaxant and analgesic. Many researchers have reported the chemical composition, antioxidant and antimicrobial activities of *Eucalyptus* species. In most species, essential oil is composed mainly of 1.8 cineole (or eucalyptol). However, the geographical distribution and variation of species greatly affect these properties, which requires extensive studies to explore the potential of this plant [8,15].

#### 3.1 Chemical constitution

The chemical composition of essential oils may vary in some situations, for example, with the region of cultivation, season and extraction method. However, the species have some characteristic constituents, which are repeated in different studies, as shown in <u>Table 1</u>. Citronellal and 1.8-cineole or eucalyptol have a concentration range between 22,30% to 78,15% and 5,90% to 89,90%, respectively, indicating that they are the most present constituents in *E. citriodora*, *E. camaldulensis* and *E. globulus* [15-25]. The studies selected for the preparation of the table did not take into account the regions, seasons or extraction methods.

Citronellal is an aldehyde and is the main component of *Cymbopogon* essential oils, *Eucalyptus citriodora*, and *Leptospermum petersonii*. Morcia C *et al.* [26] observed that citronellal was effective in reducing the in vitro growth of three species of *Fusarium*. Feyaerts AF *et al.* [27] tested 37 phytoconstituents and citronellal showed higher activity, especially against *Candida glabrata*. Tsai ML *et al.* [28] proved that citronellal exhibits antimicrobial effect against *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Escherichia coli* and *Candida albicans*. Wu Y *et al.* [29] observed that citronellal has antifungal activity against *Penicillium digitatum* and, in addition, suggested that the constituent damages the cell membrane of the fungus, increases extracellular conductivity and the release of cellular constituents.

1,8-cineole (eucalyptol) is a cyclic ether and has been reported as the main constituent of many essential oils such as *Eucalyptus camaldulensis*, *E. globulus*, *Cinnamomum longepaniculatum*, *Rosmarinus officinalis*, *Psidium pohlianum*, *P. guyanensis* and *Salvia libanotica* [30]. Sun WB *et al.* [31] reported that *Eucalyptus* showed inhibition of mycelial growth for *Fusarium oxysporum*, *F. solani* and *Cylindrocarpon destrutans*. Lee EH *et al.* [32] demonstrated that *Eucalyptus* performs anti-inflammatory function, inhibiting inflammatory signaling pathways induced by *Propionibacterium acnes*. Noumi E *et al.* [33] observed that 1,8-cineol attenuated the expression of virulence factors controlled by Quorum Sensing tested (violacein pigment production, production of elastase, protease and motility) in a dose-dependent manner for *Pseudomonas aeruginosa* and *Chromobacterium violaceum*. Şimşek M and Duman R [34] reported that cineol increased the antimicrobial activity of chlorhexidine gluconate against *Staphylococcus aureus*, *S. aureus* resistant to methicillin, *Escherichia coli*, *Klebsiella pneumoniae*, *Enterococcus faecalis* and *Candida albicans*.

Salem et al. [35] investigated the effect of the phenological stage on the yield and chemical composition of essential oils extracted from parts of *Eucalyptus globulus*. The composition analysis showed two different chemotypes depending on the growth stage which were characterized as 1,8-cineole (13.23%) at vegetative stage and p-cymene at full flowering (32.19%) and fructification (37.82%) stages. A predominance of monoterpene hydrocarbons (72.84%) during the fructification stage was detected with p-cymene (12.58%–37.82%) and  $\alpha$ -pinene (10.41%–13.39%) as the determinants of this class, the essential oil of *Eucalyptus globulus* was active against different bacterial strains, especially during the full flowering stages (MIC = 2 mg/mL) against *Bacillus cereus* and *Enterococcus faecalis*. The fruiting stage was more sensitive for *Candida albicans* than for bacteria.

In general, many studies point out that the constituents of essential oils, isolated or in combination, can exert antimicrobial activity. Among the mechanisms of action mentioned, the most accepted is that the constituents cause a rupture in the plasma membrane, affecting the proton pump and causing damage in the flow of electrons and imbalance of active transport through the membrane, besides inhibiting mitochondrial respiration of bacteria and fungi [36].

#### 3.2 Anti-Candida spp. activity

The antimicrobial activity of *E. citriodora* has been proven to be a diversity of bacterial and fungal species. Bacterial species reported are *Staphylococcus aureus*, *Methicillin-resistant S. aureus*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Haemophilus influenzae*, *Acinetobacter baumannii*, *Bacillus subtilis*, *B. cereus*, *Escherichia coli*, *Enterococcus faecalis*, *Listeria monocytogenes*, *Salmonella typhimurium*, *Agrobacterium tumefaciens*, *Dickeya solani*, *Pectobacterium atrosepticum* and *P. carotovorum*. Among the fungal species reported are *Aspergillus niger*, *A. flavus*, *A. ochraceus*, *Fusarium oxysporum*, *Penicillium funiculosum*, *P. ochrochloron*, *Rhizoctonia solani* and *Rhizopus solani* [15, 16, 37].

The activity of *E. camaldulensis* has been reported against bacteria and fungi. Among the bacterial species reported are *Staphylococcus aureus*, resistant to methicillin, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Bacillus subtilis*, *Escherichia coli* and *Acinetobacter baumannii*. Fungal species are *Aspergillus niger*, *Fusarium oxysporum*, *F. solani*, *F. verticillioides*, *F. proliferatum*, *F. subglutinans* and *Rhizopus solani*. Antiparasitic activity in *Trichomonas vaginalis* and *Plasmodium berghei* was also reported, as well as antiviral activity in the A/H1N1 virus [38-41].

Candida spp. is responsible for causing different types of infections, including those considered severe. In this sense, the evaluation of the anti-Candida spp. activity of the species of *E. citriodora*, *E. camaldulensis* and *E. globulus* is extremely important. For this, articles from the last five years were selected that prove the anti-Candida spp. activity of these species.

The activity of *E. citriodora aga*inst *Candida species* was reported by Paosen S *et al.* [45], Salem MZM *et al.* [21] and Cavalcanti AL *et al.* [46] using the broth microdilution method. The relationship of *Candida* species tested with the minimum inhibition concentration (MIC) ranges, as well as the minimum fungicide concentration (MFC) ranges are found in <u>table 2.</u>

The activity of *E. camaldulensis* against two species of *Candida* was proven by Dogan G *et al.* [47] using the disc-diffusion method, using increasing concentrations (10, 20 and 30  $\mu$ g/mL) of the essential oil extracted from the leaves and fruits of *E. camaldulensis*. The relationship of *Candida* species tested with the inhibition zone ranges is found in table 3.

The essential oil activity of *E. globulus* was analyzed using the disk-diffusion method at a concentration of 5 μL/disk. The essential oil presented a strong antifungal potential with inhibition zone of up to 25 mm. The result of the oil inhibition zone was higher in relation to the inhibition zone of the antifungals tested, Amphotericin B and Fluoroscytosine, 20 mm and 18 mm, at concentrations of 10 μg/disc and 20 μg/disc, respectively. The MIC was equivalent to 1 mg/mL. In addition, the essential oil of *E. globulus* was more sensitive to *Candida albicans* than to the bacteria also evaluated, *Staphylococcus aureus*, Methicillin-resistant *Staphylococcus aureus*, *Bacillus cereus*, *Listeria monocytogenes*, *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella pneumoniae* and *Salmonella enteridis* [35]

Thus, the essential oils of *E. citriodora*, *E. camaldulensis* and *E. globulus* present activity against different pathogens. However, further studies on anti-*Candida* spp. activity are still needed in order to obtain more consistent data, especially with *E. camaldulensis*, which only found a single study during this research.

# 3.3 Pharmacological interactions of essential oils and their phytoconstituents with antifungals against *Candida* spp.

Knezevic P *et al.* [42] determined the antimicrobial potential of two essential oils of *E. camaldulensis* against multiple clinical isolates resistant to *Acinetobacter baumanii* drugs. Antibacterial activity and synergistic effect was observed between essential oils and all antibiotics tested, ciprofloxacin, gentamicin and polymicin B. The detected MICs for the *E. camaldulensis* essential oils were in range from 0,5 to 2 μl/mL. The synergistic interaction was confirmed by time-kill curves for *E. camaldulensis* essential oil and polymyxin B combination which reduced bacterial count under detection limit very fast, after 6 h of incubation.

Silva PDC *et al.* [48] determined through *checkerboard* methodology the effect of citronellal combination with amphotercin B and ketoconazole against *Candida albicans*. The results show that the association of citronellal with ketoconazole has an additive effect against one of the strains of *C. albicans* (ICIF de 0,625) and indifferent to the other strain. While the combined activity of citronellal with amphotericin B showed an indifferent effect for all strains tested

Şimşek M and Duman R [34] investigated the efficacy of 1,8-cineole (eucalyptol), the main constituent of *E. camaldulensis*, *i*n the antimicrobial effect of chlorhexidine against some microorganisms using the *checkerboard* method. Synergistic activity has been demonstrated between chlorhexidine and 1,8-cineol against *Staphylococcus aureus*, methicillin-resistant *S. aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, *Enterococcus faecalis* and *Candida albicans*. Indifferent interactions for these compounds were demonstrated against *Pseudomonas aeruginosa*. The MIC values chlorhexidine was 2 mg/L and in combination with 1.8-cineole it was 0.5 mg/L for *C. albicans*.

Silva D *et al.* [49] evaluated the behavior of positive and negative enantiomers of  $\beta$ -citronellol, constituent of *E. citriodora*, in *Candida albicans* and *C. tropicalis strains* involved

in candidemia. Association studies have been conducted with amphotericin B using the *checkerboard* method. The two isomers showed synergistic and indifferent effect against *C. albicans* resistant to amphotericin B and *C. tropicalis*, respectively. An additive effect was also observed for the negative enantiomer against *C. albicans*. In addition, a mechanism of action assay revealed that  $\beta$ -citronellol isomers exhibited action on the fungal membrane of *Candida* spp. R-(+)- $\beta$ -citronellol and S-(-)- $\beta$ -citronellol presented a MIC 50% of 64  $\mu$ g/mL and a MFC 50% of 256  $\mu$ g/mL for *C. albicans* strains. For *C. tropicalis*, the isomers exhibited a MIC 50% of 256  $\mu$ g/mL and a MFC 50% of 1024  $\mu$ g/mL.

Tonon CC et al. [50] evaluated the antifungal activity of terpinen-4-ol associated with nystatin in biofilms of single and mixed species formed by *Candida albicans* and *Candida tropicalis*. The minimum inhibitory concentration and minimum concentration fungicide of terpinen-4-ol and nystatin were determined by the broth microdilution method, together with their synergistic activity (checkerboard method). An additive effect was observed in some concentrations of terpinen-4-ol and nystatin and also a synergistic effect at concentrations of 1.06 mg/mL of terpinen-4-ol and 0.00012 mg/mL of nystatin against *Candida albicans*. An additive effect was observed at concentrations of 1.06 mg/mL of terpinen-4-ol and 0.0003 mg/mL of nystatin against *C. tropicalis*. Terpinen-4-ol was able to reduce nystatin MIC 128 times for *C. albicans* and 64 times for *C.tropicalis*.

Salem N *et al.* [35] evaluated the synergistic effect of *E. globulus* essential oil with conventional antimicrobials against nine pathogenic bacteria and *Candida albicans*. The results of the combination of amphotericin B with the oil showed an additive effect with ICIF of 1.031. This combination showed a potential antifungal effect, the MIC value of the was 32 times lower than the oil MIC against *Candida albicans* (MIC =  $1000 \mu g/mL$ ).

During the research for the present study, no article specifically addressed the synergistic or additive effect of *E. citriodora* or *E. camaldulensis* in association with antifungals against *Candida* spp. However, studies were found on the association of the main constituents of *E. citriodora* and *E. camaldulensis* and antifungals commonly used in clinical practice. Thus, the anti-*Candida* spp. activity of essential oils has been the subject of several studies, however the evaluation of the interactions of these oils with antifungal agents are still scarce requiring greater knowledge about this as well as clarifying which mechanism of action is active.

#### **Conclusions**

Eucalyptus citriodora was effective against Candida albicans, C. tropicalis and C. krusei. Its main constituent, citronellal, was effective against C. albicans and C. glabrata and, an additive effect was observed in combination with ketoconazole against C. albicans. Eucalyptus camaldulensis was effective against C. tropicalis and C. glabrata. Its main constituent, 1,8-cineole (eucalyptol), was effective against C. albicans and synergic effect it was observed in combination with chlorhexidine against the same species. Eucalyptus globulus was effective against C. Albicans and also presented a adictive effect with amphotericin B.

The essential oils of *E. citriodora*, *E. camaldulensis* and *E. globulus*, as well as their main constituents, exhibit proven anti-*Candida* spp. activity. They also have a potential additive or synergistic effect with antimicrobials. However, further studies are still needed to consolidate the information generated in the present study, so that it can actually contribute to the therapeutic arsenal in infections caused by *Candida* spp.

#### References

- 1. Anvisa. Microbiologia Clínica para o Controle de Infecção Relacionada á Assistência á Saúde. 2013;44. Available from: https://w2.fop.unicamp.br/cibio/downloads/biosseguranca\_manutencao\_equipamentos\_labora torio\_microbiologia.pdf
- 2. Mezzari A, Fuentefria Co-autor AM. Micologia no laboratório clínico [Internet]. 2012. Available from: https://integrada.minhabiblioteca.com.br/books/9788520451762
- 3. Brooks Co-autor GF, Caroll Co-autor KC, Butel Co-autor JS, Morse Co-autor SA, Mietzner Co-autor TA. Microbiologia médica de Jawetz, Melnick e Adelberg [Internet]. 2014. Available from: https://integrada.minhabiblioteca.com.br/books/9788580553352
- 4. Zaitz C. Compendio de micologia médica [Internet]. 2010. Available from: https://integrada.minhabiblioteca.com.br/books/978-85-277-1962-9
- 5. Centers for Disease Control and Prevention (CDC) [internet]. Antifungal resistance in *Candida* [acesso em 27 jun 2020]. Available from: https://www.cdc.gov/fungal/diseases/candidiasis/antifungal-resistant.html
- 6. Ksiezopolska E, Gabaldón T. Evolutionary emergence of drug resistance in candida opportunistic pathogens. Genes (Basel). 2018;9(9).
- 7. Whaley SG, Berkow EL, Rybak JM, Nishimoto AT, Barker KS, Rogers PD. Azole antifungal resistance in Candida albicans and emerging non-albicans Candida Species. Front Microbiol. 2017;7(JAN):1–12.
- 8. D'agostino M, Tesse N, Frippiat JP, Machouart M, Debourgogne A. Essential oils and their natural active compounds presenting antifungal properties. Molecules. 2019;24(20).
- 9. Santos GCDO, Vasconcelos CC, Lopes AJO, Rocha FMG, Monteiro CDA. Candida Infections and Therapeutic Strategies: Mechanisms of Action for Traditional and Alternative Agents. 2018;9(July):1–23.
- 10. Gucwa K, Milewski S, Dymerski T, Szweda P. Investigation of the antifungal activity and mode of action of thymus vulgaris, citrus limonum, pelargonium graveolens, cinnamomum cassia, ocimum basilicum, and eugenia caryophyllus essential oils. Molecules. 2018;23(5).
- 11. Mandras N, Nostro A, Roana J, Scalas D, Banche G, Ghisetti V, et al. Liquid and vapourphase antifungal activities of essential oils against Candida albicans and non-albicans Candida. BMC Complement Altern Med [Internet]. 2016;1–7. Available from: http://dx.doi.org/10.1186/s12906-016-1316-5
- 12. Cardoso NNR, Alviano CS, Blank AF, Romanos MT V., Fonseca BB, Rozental S, et al. Synergism Effect of the Essential Oil from Ocimum basilicum var. Maria Bonita and Its Major Components with Fluconazole and Its Influence on Ergosterol Biosynthesis. Evidence-based Complement Altern Med. 2016;2016.
- 13. Bhat V, Sharma SM, Shetty V ,Shastry CS, Vaman Rao C, Shenoy S, Saha S et al. Characterization of Herbal Antifungal Agent, Origanum vulgare against Oral Candida spp.

- Isolated from Patients with Candida-Associated Denture Stomatitis: An In vitro Study. Contemp Clin Dent. 2018 Jun; 9(Suppl1): S3-S10.
- 14. de Castro RD, de Souza TMPA, Bezerra LMD, Ferreira GLS, de Brito Costa EMM, Cavalcanti AL. Antifungal activity and mode of action of thymol and its synergism with nystatin against Candida species involved with infections in the oral cavity: An in vitro study. BMC Complement Altern Med [Internet]. 2015;15(1):1–7. Available from: http://dx.doi.org/10.1186/s12906-015-0947-2
- 15. Ghaffar A, Yameen M, Kiran S, Kamal S, Jalal F, Munir B, et al. Chemical composition and in-vitro evaluation of the antimicrobial and antioxidant activities of essential oils extracted from seven eucalyptus species. Molecules. 2015;20(11):20487–98.
- 16. Chahomchuen T, Insuan O, Insuan W. Chemical profile of leaf essential oils from four Eucalyptus species from Thailand and their biological activities. Microchem J [Internet]. 2020;158(March):105248. Available from: https://doi.org/10.1016/j.microc.2020.105248
- 17. Farag NF, El-Ahmady SH, Abdelrahman EH, Naumann A, Schulz H, Azzam SM, et al. Characterization of essential oils from Myrtaceae species using ATR-IR vibrational spectroscopy coupled to chemometrics. Ind Crops Prod [Internet]. 2018;124(July):870–7. Available from: https://doi.org/10.1016/j.indcrop.2018.07.066
- 18. Gakuubi MM. Steam distillation extraction and chemical composition of essential oils of Toddalia asiatica L . and Eucalyptus. J Pharmacogn Phytochem. 2016;5(2):99–104.
- 19. Kheder DA, Al-Habib OAM, Gilardoni G, Vidari G. Components of Volatile Fractions from Eucalyptus camaldulensis Leaves from Iraqi–Kurdistan and Their Potent Spasmolytic Effects. Molecules. 2020;25(4).
- 20. Luís Â, Duarte A, Pereira L, Domingues F. Chemical Profiling and Evaluation of Antioxidant and Anti-Microbial Properties of Selected Commercial Essential Oils: A Comparative Study. Medicines. 2017;4(4):36.
- 21. Salem MZM, Elansary HO, Ali HM, El-Settawy AA, Elshikh MS, Abdel-Salam EM, et al. Bioactivity of essential oils extracted from Cupressus macrocarpa branchlets and Corymbia citriodora leaves grown in Egypt. BMC Complement Altern Med. 2018;18(1):1–7.
- 22. Tolba H, Moghrani H, Benelmouffok A, Kellou D, Maachi R. Essential oil of Algerian Eucalyptus citriodora: Chemical composition, antifungal activity. J Mycol Med. 2015;25(4):e128–33.
- 23. Pan M, Lei Q, Zang N, Zhang H. A strategy based on GC-MS/MS, UPLC-MS/MS and virtual molecular docking for analysis and prediction of bioactive compounds in Eucalyptus globulus leaves. Int J Mol Sci. 2019;20(16).
- 24. Ebani VV, Najar B, Bertelloni F, Pistelli L, Mancianti F, Nardoni S. Chemical composition and in vitro antimicrobial efficacy of sixteen essential oils against Escherichia coli and Aspergillus fumigatus isolated from poultry. Vet Sci. 2018;5(3):1–13.
- 25. Bey-Ould Si Said Z, Haddadi-Guemghar H, Boulekbache-Makhlouf L, Rigou P, Remini H, Adjaoud A, et al. Essential oils composition, antibacterial and antioxidant activities of

- hydrodistillated extract of Eucalyptus globulus fruits. Ind Crops Prod [Internet]. 2016;89:167–75. Available from: http://dx.doi.org/10.1016/j.indcrop.2016.05.018
- 26. Morcia C, Tumino G, Ghizzoni R, Bara A, Salhi N, Terzi V. In vitro evaluation of sublethal concentrations of plant-derived antifungal compounds on FUSARIA growth and mycotoxin production. Molecules. 2017;22(8).
- 27. Feyaerts AF, Mathé L, Luyten W, De Graeve S, Van Dyck K, Broekx L, et al. Essential oils and their components are a class of antifungals with potent vapour-phase-mediated anti-Candida spp. activity. Sci Rep. 2018;8(1):1–10
- 28. Tsai ML, Lin C Di, Khoo KA, Wang MY, Kuan TK, Lin WC, et al. Composition and bioactivity of essential oil from citrus grandis (L.) Osbeck 'Mato Peiyu' leaf. Molecules. 2017;22(12):1–19.
- 29. Wu Y, OuYang Q, Tao N. Plasma membrane damage contributes to antifungal activity of citronellal against Penicillium digitatum. J Food Sci Technol. 2016;53(10):3853–8.
- 30. Zomorodian K, Moein M, Pakshir K, Karami F, Sabahi Z. Chemical Composition and Antimicrobial Activities of the Essential Oil From Salvia mirzayanii Leaves. J Evidence-Based Complement Altern Med. 2017;22(4):770–6.
- 31. Sun WM, Ma YN, Yin YJ, Chen CJ, Xu FR, Dong X, et al. Effects of Essential Oils from Zingiberaceae Plants on Root-Rot Disease of Panax notoginseng. Molecules. 2018;23(5):1–11.
- 32. Lee EH, Shin JH, Kim SS, Joo JH, Choi E, Seo SR. Suppression of propionibacterium acnes-induced skin inflammation by laurus nobilis extract and its major constituent eucalyptol. Int J Mol Sci. 2019;20(14).
- 33. Noumi E, Snoussi M, Alreshidi MM, Rekha PD, Saptami K, Caputo L, et al. Chemical and biological evaluation of essential oils from cardamom species. Molecules. 2018;23(11).
- 34. Şimşek M, Duman R. Investigation of Effect of 1,8-cineole on Antimicrobial Activity of Chlorhexidine Gluconate. Pharmacognosy Res. 2017;9(3):234–7.
- 35. Salem N, Kefi S, Tabben O, Ayed A, Jallouli S, Feres N, et al. Variation in chemical composition of Eucalyptus globulus essential oil under phenological stages and evidence synergism with antimicrobial standards. Ind Crops Prod. 2018;124(June):115–25.
- 36. Leite TR, da Silva MAP, Dos Santos ACB, Coutinho HDM, Duarte AE, da Costa JGM. Antimicrobial, modulatory and chemical analysis of the oil of croton limae. Pharm Biol. 2017;55(1):2015–9.
- 37. Miguel M, Gago C, Antunes M, Lagoas S, Faleiro M, Megías C, et al. Antibacterial, Antioxidant, and Antiproliferative Activities of Corymbia citriodora and the Essential Oils of Eight Eucalyptus Species. Medicines. 2018;5(3):61.
- 38. Anigbo AA, Avwioroko OJ, Cholu CO. Phytochemical Constituents, Antimalarial Efficacy, and Protective Effect of Eucalyptus camaldulensis Aqueous Leaf Extract in Plasmodium berghei-Infected Mice. Prev Nutr Food Sci. 2020;25(1):58–64.

- 39. Aslani A, Asghari G, Darani HY, Ghanadian M, Hosseini F. Design, formulation, and physicochemical evaluation of vaginal cream containing Eucalyptus camaldulensis, Viola odorata, and Mentha piperita extracts for prevention and treatment of trichomoniasis. Int J Prev Med [Internet]. 2019 Jan 1 [cited 2020 Jul 28];10(1). Available from: /pmc/articles/PMC6826777/?report=abstract
- 40. Butts A, Palmer GE, Rogers PD. Antifungal adjuvants: Preserving and extending the antifungal arsenal. Virulence [Internet]. 2017;8(2):198–210. Available from: http://dx.doi.org/10.1080/21505594.2016.1216283
- 41. Chaves TP, Pinheiro REE, Melo ES, Soares MJ dos S, Souza JSN, Andrade TB de, et al. Essential oil of Eucalyptus camaldulensis Dehn potentiates β-lactam activity against Staphylococcus aureus and Escherichia coli resistant strains. Ind Crops Prod [Internet]. 2018;112(June 2017):70–4. Available from: https://doi.org/10.1016/j.indcrop.2017.10.048
- 42. Knezevic P, Aleksic V, Simin N, Svircev E, Petrovic A, Mimica-Dukic N. Antimicrobial activity of Eucalyptus camaldulensis essential oils and their interactions with conventional antimicrobial agents against multi-drug resistant Acinetobacter baumannii. J Ethnopharmacol [Internet]. 2016;178:125–36. Available from: http://dx.doi.org/10.1016/j.jep.2015.12.008
- 43. Sadatrasul MS, Fiezi N, Ghasemian N, Shenagari M. Oil-in-water emulsion formulated with eucalyptus leaves extract inhibit influenza virus binding and replication in vitro. 2017;3(November):899–907.
- 44. Upreti A, Byanju B, Fuyal M, Chhetri A, Pandey P, Ranjitkar R, et al. Evaluation of α-amylase, lipase inhibition and in-vivo pharmacological activities of Eucalyptus camaladulensis Dehnh leaf extract. J Tradit Complement Med [Internet]. 2019;9(4):312–8. Available from: https://doi.org/10.1016/j.jtcme.2018.07.001
- 45. Paosen S, Jindapol S, Soontarach R, Voravuthikunchai SP. Eucalyptus citriodora leaf extract-mediated biosynthesis of silver nanoparticles: broad antimicrobial spectrum and mechanisms of action against hospital-acquired pathogens. Apmis. 2019;127(12):764–78.
- 46. Cavalcanti AL, Aguiar YPC, Santos FG Dos, Cavalcanti AFC, De Castro RD. Susceptibility of candida albicans and candida non-albicans strains to essential oils. Biomed Pharmacol J. 2017;10(3):1101–7.
- 47. Dogan G, Kara N, Bagci E, Gur S. Chemical composition and biological activities of leaf and fruit essential oils from Eucalyptus camaldulensis. Zeitschrift fur Naturforsch Sect C J Biosci. 2017;72(11–12):483–9.
- 48. Silva PDC, Santos BLC dos, Soares GL, Oliveira WA de. Anti-Candida spp. albicans activity of the association of citronelal with anfotericin B or with cetoconazole. Period Tche Quim. 2019;16(31):250–7.
- 49. Silva D, Diniz-neto H, Silva-neta M, Silva S, Andrade-j F, Leite M, et al. (R)-(+)-β-Citronellol and (S)-(-)-β-Citronellol in Combination with Amphotericin B against Candida Spp. Int J Mol Sci Artic. 2020.

50. Tonon CC, Francisconi RS, Bordini EAF, Huacho PMM, Sardi J de CO, Spolidorio DMP. Interactions between terpinen-4-ol and nystatin on biofilm of Candida albicans and Candida tropicalis. Braz Dent J. 2018;29(4):359–67.

## **Tables**

**Table 1.** Chemical composition of *E. citriodora*, *E. camaldulensis* and *E. globulus*.

SPECIES	CONSTITUENTS	CONCENTRATION RANGE	
	citronellal	22,30% - 78,15%	
Eucalyptus citriodora	citronellol	5,55% - 20,00%	
	citronellol acetate	1,33% - 12,30%	
	isopulegol	1,12% - 7,60%	
	α-pineno	1,15% - 3,60%	
	eucalyptol	2,00% - 2,50%	
	1.8-cineole	5,90% - 62,70%	
	p-cymene	6,70% - 35,70%	
Eucalyptus camaldulensis	terpinene	10,70% - 22,04%	
	α-pinene	3,03% - 15,60%	
	terpinen-4-ol	2,00% - 5,30%	
	α-terpineol	2,85% - 4,40%	
Eucalyptus globulus	1.8-cineole	13,23 – 89,80%	
	α-pinene	2,00 -16,06%	
	Aromadendrene	0,57 -19,70%	
	o-Cymene	0,50 – 2,35%	
	D-Limonene	0,30 - 2,59%	
	Camphene	0,19 - 2,43%	

 Table 2. Anti-Candida spp. activity of E. citriodora essential oil.

E. citriodora	Candida spp.	MIC	MFC RANGE
	C. albicans	0,02 μg/mL - 0,25 mg/mL	0,09 μg/mL - 0,52 mg/mL
	C. krusei	5 μg/mL	5 μg/mL
	C. tropicalis	2,5 μg/mL	10 μg/mL

 Table 3. Anti-Candida spp. activity of E. camaldulensis essential oil.

		INHIBITION ZONE	
	Candida spp.	Leaves	Fruits
E. camaldulensis	C. tropicalis	18 - 22 mm	12 - 18 mm
	C. glabrata	19 - 23 mm	13 - 20 mm

# ATTACHMENT I: Instructions For Authors - "Journal of Innovations in Pharmaceutical and Biological Sciences"

**Manuscript Preparation:** The manuscript should be typed single-spaced on A4  $(8.5" \times 11")$  paper size with 1 inch margins on all sides. Times New Roman font 12 should be used. Manuscript should be arranged in the following order: Title, Abstract, Keywords, Introduction, Materials and Methods, Results, Discussion, Conclusion, Acknowledgement, References, Figures and Tables.

**Title Page:** The title page should contain a clear, concise and informative title of the article followed by the names and affiliations of the authors. The affiliation should comprise the department, institution, city, and state (or nation) and should be typed as a footnote to the author's name. The Corresponding Author must indicate his or her complete mailing address, office/cellular phone number, fax number, and email address at the lower left of the Title Page.

**Abstract:** The abstract should start on a new page after the title page and should not be more than 250 words and should contain objectives, material and methods, Results and Conclusions. Reviews and mini reviews also require an abstract.

**Keywords:** Following the abstract, provide a maximum of 5-6 keywords, which reflect the content of the study.

**Introduction:** This should be brief and indicates aim of the study and the essential back ground information. The introduction should not be an extensive literature review although it should provide sufficient background information for the reader to understand and evaluate the results of the present study without referring to previous publications on the same topic.

**Material and methods:** Please provide concise but complete information about the material and the analytical, statistical and experimental procedures used. This part should be as clear as possible to enable other scientists to repeat the research presented. In case of animal/human experiments or clinical trials authors must give the details of ethical approval.

**Result and Discussion:** Data acquired from the research with appropriate statistical analysis described in the methods section should be included in this section. In this part, the same data/information given in a table must not be repeated in a figure, or vice versa. Tables and figures should be self explanatory and it is not acceptable to repeat extensively the numerals from tables into text and give lengthy and unnecessary explanations of the Tables and figures. Discussion should relate the results to current understanding of the scientific problems being investigated in the field.

**Conclusion:** A short, paragraph summarizing the most important finding(s) of the research is required.

**Acknowledgement:** All acknowledgments (if any) should be included at the very end of the paper before the references and may include supporting grants, presentations, and so forth.

**References:** Should be numbered consecutively in the order in which they are first mentioned in the text (not in alphabetic order). Indicate references by number(s) in square brackets [Reference no] in line with the text References cited only in tables or figure legends should be numbered in accordance with the sequence established by the first identification in the text of the particular table or figure.

**Journal Articles:** Shashi A, Jain SK and Pandey M: In-vitro evaluation of antilthiatic activity of seeds of Dolichos biflorus and roots of Asparagus racemosus. International Journal of Plant Sciences 2008; 1:67-71.

**Tables & Figures:** Tables and figures should not be embedded in the text, but should be included at the end of the manuscript on separate pages. Tables should be created with a word processor and cited consecutively in the text. Number tables and figures consecutively in accordance with their appearance in the text. Place footnotes to tables below the table and give proper heading on table.

**Abbreviations:** Standard abbreviations should be used throughout the manuscript. Use of nonstandard abbreviations can be confusing to readers.