NITROUS OXIDE AND METHANE FLUXES IN SOUTH BRAZILIAN GLEY SOL AS AFFECTED BY NITROGEN FERTILIZERS(1)

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SUMMARY

Nitrogen fertilizers increase the nitrous oxide (N2O) emission and can reduce the methane (CH4) oxidation from agricultural soils. However, the magnitude of this effect is unknown in Southern Brazilian edaphoclimatic conditions, as well as the potential of different sources of mineral N fertilizers in such an effect. The aim of this study was to investigate the effects of different mineral N sources (urea, ammonium sulphate, calcium nitrate, ammonium nitrate, Uran, controlled-release N fertilizer, and urea with urease inhibitor) on N2O and CH4 fluxes from Gleysol in the South of Brazil (Porto Alegre, RS), in comparison to a control treatment without a N application. The experiment was arranged in a randomized block with three replications, and the N fertilizer was applied to corn at the V5 growth stage. Air samples were collected from a static chambers for 15 days after the N application and the N2O and CH4 concentration were determined by gas chromatography. The topmost emissions occurred three days after the N fertilizer application and ranged from 187.8 to 858.2 μg m⁻² h⁻¹ N. The greatest emissions were observed for N-nitric based fertilizers, while N sources with a urease inhibitor and controlled release N presented the smallest values and the N-ammonium and amidic were intermediate. This peak of N2O emissions was related to soil NO3-N (R² = 0.56, p < 0.08) when the soil water-filled pore space was up to 70 % and it indicated that N2O was predominantly produced by a denitrification process in the soil. Soil CH4 fluxes ranged from -30.1 μg m⁻² h⁻¹ C (absorption) to +32.5 μg m⁻² h⁻¹ C (emission), and the accumulated emission in the period was related to the soil NH4+-N concentration (R² = 0.82, p < 0.001), probably due to enzymatic competition between nitrification and metanotrophy processes. Despite both of the gas fluxes being

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affected by N fertilizers, in the average of the treatments, the impact on CH₄ emission (0.2 kg ha⁻¹ equivalent CO₂-C) was a hundredfold minor than for N₂O (132.8 kg ha⁻¹ equivalent CO₂-C). Accounting for the N₄O and CH₄ emissions plus energetic costs of N fertilizers of 1.3 kg CO₂-C kg⁻¹ N regarding the manufacture, transport and application, we estimated an environmental impact of N sources ranging from 220.4 to 664.5 kg ha⁻¹ CO₂-C, which can only be partially offset by C sequestration in the soil, as no study in South Brazil reported an annual net soil C accumulation rate larger than 160 kg ha⁻¹ C due to N fertilization. The N₂O mitigation can be obtained by the replacement of N-nitric sources by ammonium and amidic fertilizers. Controlled release N fertilizers and urea with urease inhibitor are also potential alternatives to N₂O emission mitigation to atmospheric and systematic studies are necessary to quantify their potential in Brazilian agroecosystems.

Index terms: nitrogen fertilization, environmental impacts, agriculture, greenhouse effect.

RESUMO: FLUXOS DE ÓXIDO NITROSO E DE METANO EM GLEISSOLO INFLUENCIADOS PELA APLICAÇÃO DE FERTILIZANTES NITROGENADOS NO SUL DO BRASIL

Fertilizantes nitrogenados incrementam os fluxos de óxido nitroso (N₂O) e podem deprimir a oxidação de metano (CH₄) em solos agrícolas. Entretanto, não existem resultados da magnitude desses efeitos nas condições edafoclimáticas do Sul do Brasil, tampouco do potencial de algumas fontes de N em mitigar esses efeitos. O presente estudo objetivou avaliar o impacto da aplicação de fertilizantes nitrogenados (ureia, sulfato de amônio, nitrito de cálcio, nitrito de amônio, UraN, N de liberação lenta e ureia com inibidor de urease) nos fluxos de N₂O e CH₄ em um Gleissolo no Sul do Brasil (Porto Alegre, RS), em comparação a um tratamento controle sem aplicação de N. O experimento seguiu um delineamento de blocos ao acaso, com três repetições, e os fertilizantes foram aplicados, em cobertura, numa dose única de 150 kg ha⁻¹ N, no estádio V5 da cultura do milho. A avaliação dos gases foi feita utilizando-se o método da câmara estática, nos 15 dias que sucederam a aplicação de N, e a análise das concentrações de N₂O e CH₄ nas amostras de ar foi realizada por meio de cromatografia gasosa. O pico de emissão de N₂O ocorreu no terceiro dia após a aplicação dos fertilizantes nitrogenados e a sua intensidade variou de 187,8 a 8.587,4 μg m⁻² h⁻¹ N, destacando-se as fontes nítricas com as maiores emissões, as fontes amoniacais e amídicas com emissões intermediárias e os fertilizantes de liberação lenta e com inibidor de urease com as menores emissões. As emissões no terceiro dia tiveram relação direta com os teores de N-NO₃ do solo (R² = 0,56; p < 0,08) e ocorreram quando este apresentava valores de porosidade preenchida por água (PPA) maiores que 70 %, o que indica que a desnitrificação foi o processo predominante na produção de N₂O. Os fluxos de CH₄ do solo variaram de -30,1 μg m⁻² h⁻¹ C (absorção) a +32,5 μg m⁻² h⁻¹ C (emissão), e a emissão acumulada desse gás teve relação direta com os teores de NH₄⁺ no solo (R² = 0,82; p < 0,001), possivelmente pela competição enzimática entre os processos de nitrificação e de metanotrofia. Apesar de os fluxos de ambos os gases terem sido alterados pela aplicação dos fertilizantes nitrogenados, na média dos tratamentos, o impacto das emissões de CH₄ (0,2 kg ha⁻¹ C-CO₂ equivalente) foi centenas de vezes menor que o verificado para as emissões de N₂O (132,8 kg ha⁻¹ C-CO₂ equivalente). Considerando as emissões desses gases no solo fertilizado e o custo médio de 1,3 kg C-CO₂ kg⁻¹ N referente à produção, transporte e aplicação do fertilizante, o impacto ambiental dos fertilizantes nitrogenados variou de 220,4 a 664,5 kg ha⁻¹ C-CO₂, o qual pode ser apenas parcialmente contrabalanceado pelo acúmulo de C na matéria orgânica do solo, pois nenhum estudo realizado no Sul do Brasil evidenciou taxa anual de acúmulo de C no solo, decorrente da adubação nitrogenada, maior que 160 kg ha⁻¹ C. A redução das emissões de N₂O do solo e, portanto, do impacto ambiental pode ser obtida pelo uso de fontes amoniacais e amídicas em detrimento de fontes nítricas. Os fertilizantes de liberação lenta e com inibidores de urease também são alternativas potenciais visando à mitigação das emissões de N₂O para atmosfera, e esforços deverão ser empreendidos numa avaliação sistemática desse potencial em agroecossistemas brasileiros.

Termos de indexação: adubação nitrogenada, impacto ambiental, agricultura, efeito estufa.
INTRODUCTION

Mineral N application to the soil is a key practice for achieving high crop yield in non leguminous species such as corn, sugarcane and winter cereal crops (Amado et al., 2002; Conti, 2007; Fontoura & Bayer, 2009). The Brazilian consumption of N fertilizer has increased at 5 % per year in the last decade, surpassing two million tons of annual demand (Lopes & Bastos, 2007). According to the national inventory of greenhouse gases (GHG) emission, N fertilization is one of the main sources of nitrous oxide (N₂O) emission from agriculture in the country (Brasil, 2006) and, in addition, studies have evidenced depressive effects of such practice on methane (CH₄) oxidation in agricultural soils (Mosier et al., 1991; Hütsch, 2001). Despite the tendency of increasing the use of N fertilizers, however, few efforts have been made aiming to evaluate the magnitude of these effects and the potential of different sources of N fertilizers in minimizing the GHG emissions, particularly the N₂O, in the Brazilian agricultural systems (Steudler et al., 2002; Carvalho et al., 2006).

The N fertilization increases the mineral N availability in the soil (NO₃⁻-N + NH₄⁺-N), which is a substrate for the nitrification and denitrification processes, which in turn are responsible for the N₂O production in the soil (Khalil et al., 2004). The denitrification is the direct source of N₂O from nitric N-based fertilizers, while both processes of nitrification and denitrification can be involved in N₂O emissions from amidic and ammoniacal N sources (Liu et al., 2007). Although both processes can contribute to the N₂O emission from nitric N-based fertilizers, while both processes of nitrification and denitrification can be involved in N₂O emissions from amidic and ammoniacal N sources, several studies suggested that denitrification is the dominant process from soils in which the water-filled pore space (WFPS) is larger than 70 % (Bateman & Baggs, 2005; Liu et al., 2007). Taking such results into account, it is reasonable to infer that the application of nitric N-based sources might promote greater N₂O emissions than amidic and ammoniacal sources in poorly aerated soils (> 70 % WFPS).

Oppositely, however, a decline in the potential of CH₄ oxidation has been attributed to the amidic and ammoniacal sources due to their short and long term effects on soil methanotrophy rates (Hütsch, 2001; Kravchenko et al., 2002). Nitric-based fertilizers usually have a small effect on the soil CH₄ oxidation capacity. However, if applied at high rates and depending upon the accompanying cation, they can significantly enhance the CH₄ emission in consequence of the injurious effect of salinity on methanotrophic bacteria and cause the rise of NH₄⁺ concentration in soil solution from its displacement from exchange sites (Hütsch, 2001).

Besides the outcomes from amidic/ammoniacal and nitric N sources regarding soil N₂O and CH₄ emissions, the adoption of microbial activity inhibitors and controlled-release fertilizers assume a prominent role in scenarios of mitigation of GHG emissions. Such inputs have markedly decreased the N losses by NH₃ volatilization (Conti, 2007; Fontoura et al., 2010). Moreover, their delaying effect on the N fertilizer transformation into NH₄⁺-N and, consequently, into NO₃⁻-N, can possibly cause a decline in N₂O emissions to atmosphere (Zaman et al., 2008; Snyder et al., 2009).

The study is part of an endeavor of identifying strategies of mitigating GHG emission in Brazilian agricultural systems and aimed to evaluate the impact of different N fertilizers on N₂O and CH₄ emissions in a South Brazilian Gleysol.

MATERIAL AND METHODS

Soil and climate characteristics

The study was performed on the experimental area of the Agronomy Faculty, at the Federal University of the State of Rio Grande do Sul - UFRGS, in Porto Alegre, RS (30 ° 04 ' S and 51 ° 08 ' W). The local climate is subtropical humid, with warm summers and cold and rainy winters (Cfa according to Koöpen classification). The mean annual temperature is 19.4 °C and January is the hottest month – with a mean of 25 °C. The mean annual rainfall is 1,299 mm regularly distributed over the year. The soil is classified as Gleysol (FAO) and has 260 g kg⁻¹ clay and 310 g kg⁻¹ sand (0 to 0.20 m). Some chemical attributes of the soil before the onset of the experiment are reported in table 1.

Experimental design and treatments

The experiment was carried out on an area in which, for about ten years, it has been previously cultivated with corn (Zea mays L.) under no tillage system in

Table 1. Soil attributes in a Gleysol at the beginning of the experiment

<table>
<thead>
<tr>
<th>Layer</th>
<th>C g kg⁻¹</th>
<th>pH</th>
<th>P cm o.l. kg⁻¹</th>
<th>K cm o.l. kg⁻¹</th>
<th>Ca²⁺ cm o.l. kg⁻¹</th>
<th>Mg²⁺ cm o.l. kg⁻¹</th>
<th>Al³⁺ cm o.l. kg⁻¹</th>
<th>CTC ef. cm o.l. kg⁻¹</th>
<th>Density Mg m⁻³</th>
</tr>
</thead>
<tbody>
<tr>
<td>m</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0 to 0.05</td>
<td>25.3</td>
<td>4.9</td>
<td>16.0</td>
<td>105</td>
<td>3.4</td>
<td>1.5</td>
<td>0.5</td>
<td>5.7</td>
<td>1.35</td>
</tr>
<tr>
<td>0.05 to 0.10</td>
<td>16.0</td>
<td>5.0</td>
<td>7.0</td>
<td>66</td>
<td>3.1</td>
<td>1.3</td>
<td>0.5</td>
<td>5.1</td>
<td>1.47</td>
</tr>
</tbody>
</table>
the summer and ryegrass in the winter (*Lolium multiflorum* L.), propagated by natural re-seeding. In the occasion of the experiment establishment, in October 2007, the ryegrass dry mass (~3 Mg ha⁻¹ DM) was desiccated with glyphosate-based herbicide (3.5 L ha⁻¹). Corn was manually sowed at 15 days after herbicide application, with an inter row distance of 0.7 m and number of seeds for a final population of about 55,000 plants ha⁻¹. Sowing fertilization was applied for all treatments and consisted of 30 kg ha⁻¹ of N, 60 of P₂O₅ and 60 of K₂O, by applying urea, triple super phosphate and potassium chloride, respectively. The sidedress fertilization (150 kg ha⁻¹ N) was performed in a single application when corn plants reached the V5 stage. The mineral N sources [ammonium nitrate (NH₄NO₃); calcium nitrate (Ca(NO₃)₂); ammonium sulphate [(NH₄)₂SO₄]; urea (CO(NH₂)₂); urea with urease inhibitor; controlled-release N; and Uran, a liquid formula of N, obtained from a physical mix of urea and ammonium nitrate], in addition to the control (without sidedress N fertilization), were applied according to a randomized block design, with three replicates. The corresponding fertilizer was applied on the surface, without incorporation, at about 0.10 m from the corn row. A proportional rate of each N fertilizer was applied inside the bases area (placed in the inter row region) where the N₂O and CH₄ fluxes were evaluated. The controlled-release N source (Sulfammo®) is composed by N molecules coated by an organo-mineral double layer that slows down the N dissolution by preventing the water entrance into its interior. The urea with urease inhibitor (Super N®) is obtained from the NBPT (N-(n-butyl) thiophosphoric triamide) addition to the urea. The NBPT occupies the active site of urease, postponing the enzyme action for three to seven days. The relative composition of N chemical species in each mineral N fertilizer is reported in table 2.

**Air sampling and soil N₂O and CH₄ analysis**

Soil fluxes of N₂O and CH₄ were evaluated by using the static chamber method (Jantalia et al., 2008). In the inter row spacing, three aluminum bases per treatment (constituting three replicates) were inserted into the soil at a depth of 5 cm. The bases were covered with plastic film when the fertilizers were applied to the area. Subsequently, the plastic was removed and each base received the corresponding N source proportionally to the base area (346.5 cm²). The PVC chambers (20 cm height and 25 cm diameter) were allocated on a canal, in the top of each base, which was filled with water to hermetically close the chamber volume during the air sampling events. In the top, each chamber contained a triple valve for fastening the sampling syringes, a digital thermometer with an outside display for monitoring the inner temperature of the chamber and a hole 1 cm in diameter for equilibrating the pressure during the placement of the chambers on the canal. This hole was kept closed during the air sampling by using a rubber septum.

Air sampling and N₂O and CH₄ analysis were performed at 0, 1, 3, 4, 5, 6, 7, 9, 12 and 15 days after the N sidedress application in corn. On the second day after application, air was not sampled due to rain occurrence at the moment of the sampling (28 mm). The sampling events always started at 9 a.m., assuming that the emissions at that time represent the approximate daily mean of GHG emissions (Jantalia et al., 2008). Air samples from the interior of each chamber were taken at 0 (zero), 15, 30 and 45 min after closing chambers, using polypropylene syringes (20 mL) containing a triple Luer lock valve in its tip. Immediately before each sampling time, the inner atmosphere was homogenized through an internal fan for 30 s. The inner temperature of the chamber was monitored at every sampling time through the digital thermometer allocated in one replicate of each treatment. Additional information regarding the material and the sampling procedure are available in Gomes et al. (2009) and Zanatta (2009).

The syringes containing the air samples were refrigerated in thermic boxes at low temperatures and were transported to the lab of Environmental Biochemistry (UFRGS), where they were analyzed by gas chromatography up to 36 h after sampling. The

<table>
<thead>
<tr>
<th>Fertilizer</th>
<th>% total N(¹)</th>
<th>kg N of each chemical species(²)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>NO₃⁻</td>
</tr>
<tr>
<td>Urea</td>
<td>45</td>
<td>-</td>
</tr>
<tr>
<td>Ammonium sulphate</td>
<td>26</td>
<td>-</td>
</tr>
<tr>
<td>Calcium nitrate</td>
<td>15</td>
<td>140</td>
</tr>
<tr>
<td>Ammonium nitrate</td>
<td>34</td>
<td>75</td>
</tr>
<tr>
<td>Uran</td>
<td>32</td>
<td>42</td>
</tr>
<tr>
<td>Controlled-release N(³)</td>
<td>23</td>
<td>26</td>
</tr>
<tr>
<td>Urea with urease inhibitor(⁴)</td>
<td>45</td>
<td>-</td>
</tr>
</tbody>
</table>

(¹) Brand name: sulfammo. (²) Brand name: super N. (³) Values based on the manufacturer warranty.
equipment was a Shimadzu GC 2014 model “Greenhouse”, equipped with three packed columns working at 70 °C, N₂ as a carrier gas at a flow of 26 mL min⁻¹, injector with loop for direct sampling of 1 mL and temperature set at 250 °C, electron capture detector (ECD) at 325 °C for N₂O detection and flame ionization detector (FID) at 250 °C for CH₄.

The N₂O and CH₄ fluxes were estimated based on the following equation.

\[ f = \frac{ΔQ}{Δt} \frac{PV}{RT} \frac{1}{A} \]

where, \( f \) is the methane or nitrous oxide flux (μg m⁻² h⁻¹ N₂O or CH₄), \( Q \) is the quantity of each gas in the chamber at the sampling moment (μg N₂O or CH₄/ chamber), \( P \) is the atmospheric pressure (atm) in the chamber, assumed as being 1 atm, \( V \) is the chamber volume (L), \( R \) is the ideal gases constant (0.08205 atm L mol⁻¹ K⁻¹), \( T \) is the inner temperature of the chamber at the sampling moment (°K) and \( A \) is the chamber area (m²). The rate of gas concentration increase in the chamber was estimated by using the angular coefficient obtained from the relationship between gas concentration and time. The daily emissions were estimated from the rates of fluxes and the total emission for the period (15 days) they were emissions were estimated from the rates of fluxes and the total emission for the period (15 days) they were calculated from the integration of the daily N₂O and CH₄ emission from soil versus time (Gomes et al., 2009). The accumulated CH₄ and N₂O emission for the experiment period was converted to quantities of equivalent CO₂-C per hectare by taking into account the global warming potential of each gas in comparison to CO₂ (23 times greater for CH₄ and 296 greater for N₂O) (Mosier et al., 2006).

**Soil and climate variables**

Some soil variables were evaluated in each air sampling event in order to identify driving variables for N₂O and CH₄ emissions. At the 0 to 0.10 m, the gravimetric water content, pH-H₂O and the NO₃⁻--N and NH₄⁺--N contents were analyzed in compound samples (5 sub-samples/plot) according to methods described by Tedesco et al. (1995). The water-filled pore space (WFPS) was estimated from the gravimetric water content and the soil bulk density (Gomes et al., 2009), which was analyzed by using the core method at the end of the period of 15 days of evaluation and whose results are reported in table 1. Local data of rainfall for the evaluated period were obtained from an Automatic Meteorological Station of the National Institute of Meteorology, located about 5 km from the experiment area.

**Statistical analysis**

The effects of N fertilizers on accumulated CH₄ and N₂O emission and on the N₂O emission peak were evaluated by orthogonal contrasts (p < 10 %). The identification of the soil driving variables for CH₄ and N₂O emission was performed by evaluating the significance of determinant coefficients (R²) of polynomial regression and multiple linear regressions.

**RESULTS AND DISCUSSION**

**Soil N₂O fluxes**

The N₂O emission from soil was remarkably affected by N fertilization (Figure 1b, Tables 3 and 4). In comparison to the unfertilized soil (control), N fertilization raised the rates of N₂O emission from soil at about 4 to 400 times, reaching the upmost rates at the third day after fertilization – which coincided with WFPS values larger than 70 % (Figure 1a) and with the largest NO₃⁻--N and NH₄⁺--N contents in the soil (Figure 2a-b). Despite the intensity, the N₂O peak of emission dropped sharply, reaching rates noticeably smaller at only one day subsequent to the peak of emission (mean for N-fertilized soils = 71.1 μg m⁻² h⁻¹ N), but sustaining greater rates than the mean of the unfertilized soil (15.0 μg m⁻² h⁻¹ N) up to the 12th day (see detail in Figure 1b).

Several studies have depicted the occurrence of peaks in N₂O emission following N fertilization (Clayton et al., 1997; Dobbie & Smith, 2003; Venterea et al., 2005). In Brazil, a pioneer study performed in the Northern region documented increases of N₂O emission rates at 15 to 18 times in an Ultisol after NO₃⁻--N based fertilization (Steudler et al., 2002). However, in an Oxisol of the Cerrado region, a recent study did not evidence increases in N₂O emissions after the application of 60 kg ha⁻¹ of urea, which was attributed to the low water content in the soil at that moment (Carvalho et al., 2006), possibly enhanced by the fast drainage of that soil.

The sources of N fertilizers affected, in distinct magnitudes, the peak of N₂O emission from the soil (Figure 1b). The application of nitric forms promoted the largest N₂O emission rates (8,587.4 and 3,485.8 μg m⁻² h⁻¹ N for ammonium nitrate and calcium nitrate, respectively). The N₂O emission for ammonium sulphate- and urea-amended soils peaked at 1,435.3 and 859.1 μg m⁻² h⁻¹ N, respectively (Figure 1b), while the smallest peaks of emission rates were found from soils amended with controlled-release N (187.8 μg m⁻² h⁻¹ N) and with urea with urease inhibitor (346.4 μg m⁻² h⁻¹ N). The Uran fertilizer, composed of a mix of urea and ammonium nitrate, fostered peaks of intermediate intensity in comparison to the application of these sources individually (2,050.7 μg m⁻² h⁻¹ N). Although statistically similar, the smaller N₂O emission due to urea amendment than ammonium sulphate might be due to the possibly larger loss of nitrogen by volatilization, which would diminish the NH₄⁺--N and NO₃⁻--N generation and their outcomes on N₂O emission.
Larger \( \text{N}_2\text{O} \) emission rates have been described following ammonium nitrate application than for other mineral N sources (Clayton et al., 1997; Maggiotto et al., 2000; Dobbie & Smith, 2003; Venterea et al., 2005). The magnitude of the emissions in these reports, however, were usually smaller than the emissions herein depicted, though large rates – similar to the current study – were already documented by Maggiotto et al. (2000) for soils under pasture with ryegrass in raining years (7.528 \( \mu \text{g m}^{-2} \text{h}^{-1} \text{N} \)). In the current study, the large \( \text{N}_2\text{O} \) emissions subsequent to nitric-N fertilization were likely exacerbated by associated effects of rain occurrence in the beginning of the evaluation period, poor drainage of the soil and the elevated water table. Such combination of factors, that nourished anoxic conditions due to the large values of WFPS, associated to the great soil contents of \( \text{NO}_3^-\text{N} \) remarkably accentuated the N loss as \( \text{N}_2\text{O} \). In other types of soils with similar physical and chemical attributes and under analogous climatic conditions, it is possible that the \( \text{N}_2\text{O} \) emissions would be smaller than those obtained in this study. The particular features of the Gleysol regarding the drainage and the proximity to the water table determines the presence of a distinguished microbial population, which is tipical for the frequent anoxic and reduced conditions of these soils and would not dominate in soils under better drainage status.

![Figure 1](image.png)

Figure 1. Rainfall, water-filled pore space (WFPS) at the 0 to 0.10 m soil layer (a) and \( \text{N}_2\text{O} \) emission (\( \mu \text{g m}^{-2} \text{h}^{-1} \text{N} \)) from soil (b) for the 15 days period after application of the nitrogen fertilizers to a Gleysol in the South of Brazil. Vertical bars mean standard deviation of the mean (\( n=3 \)).

<table>
<thead>
<tr>
<th>Orthogonal contrasts</th>
<th>Peak 3rd day ( \text{N}_2\text{O} )</th>
<th>Accumulated emission ( \text{N}_2\text{O} )</th>
<th>( \text{CH}_4 )</th>
</tr>
</thead>
<tbody>
<tr>
<td>No N appl. x With N appl. (all sources)</td>
<td>0.06</td>
<td>0.07</td>
<td>0.01</td>
</tr>
<tr>
<td>Urea x Ammonium sulphate</td>
<td>0.19</td>
<td>0.30</td>
<td>0.09</td>
</tr>
<tr>
<td>Urea x Calcium nitrate+Ammonium nitrate</td>
<td>0.03</td>
<td>0.03</td>
<td>0.43</td>
</tr>
<tr>
<td>Calcium nitrate x Ammonium nitrate</td>
<td>0.10</td>
<td>0.09</td>
<td>0.50</td>
</tr>
<tr>
<td>Ammonium nitrate x all other sources</td>
<td>0.01</td>
<td>0.01</td>
<td>0.28</td>
</tr>
<tr>
<td>Urea+Ammonium sulphate x Urease inhibitor and Controlled-release N</td>
<td>0.02</td>
<td>0.02</td>
<td>0.54</td>
</tr>
</tbody>
</table>

Larger \( \text{N}_2\text{O} \) emission rates have been described following ammonium nitrate application than for other mineral N sources (Clayton et al., 1997; Maggiotto et al., 2000; Dobbie & Smith, 2003; Venterea et al., 2005). The magnitude of the emissions in these reports, however, were usually smaller than the emissions herein depicted, though large rates – similar to the current study – were already documented by Maggiotto et al. (2000) for soils under pasture with ryegrass in raining years (7.528 \( \mu \text{g m}^{-2} \text{h}^{-1} \text{N} \)). In the current study, the large \( \text{N}_2\text{O} \) emissions subsequent to nitric-N fertilization were likely exacerbated by associated effects of rain occurrence in the beginning of the evaluation period, poor drainage of the soil and the elevated water table. Such combination of factors, that nourished anoxic conditions due to the large values of WFPS, associated to the great soil contents of \( \text{NO}_3^-\text{N} \) remarkably accentuated the N loss as \( \text{N}_2\text{O} \). In other types of soils with similar physical and chemical attributes and under analogous climatic conditions, it is possible that the \( \text{N}_2\text{O} \) emissions would be smaller than those obtained in this study. The particular features of the Gleysol regarding the drainage and the proximity to the water table determines the presence of a distinguished microbial population, which is tipical for the frequent anoxic and reduced conditions of these soils and would not dominate in soils under better drainage status. For
Table 4. Accumulated N₂O and CH₄ emissions from soil in the 15 day period after nitrogen fertilizer application, their correspondent values in equivalent CO₂-C, costs in CO₂-C of the fertilizer and total greenhouse gases emission (N₂O+CH₄+custos C-CO₂) as a consequence of mineral nitrogen fertilization

<table>
<thead>
<tr>
<th>Fertilizer</th>
<th>N₂O</th>
<th>CH₄</th>
<th>Cost(1) C-CO₂</th>
<th>Total emission</th>
</tr>
</thead>
<tbody>
<tr>
<td>No mineral N</td>
<td>&lt;0.01±0.01</td>
<td>6.9±1.1</td>
<td>-45.1±10.9</td>
<td>-377.1±7.1</td>
</tr>
<tr>
<td>Urea</td>
<td>0.48±0.21</td>
<td>67.4±15.3</td>
<td>26.3±19.1</td>
<td>-220.0±63.7</td>
</tr>
<tr>
<td>Urea+Urease Inh.</td>
<td>0.33±0.23</td>
<td>48.3±16.7</td>
<td>12.4±1.2</td>
<td>-104.0±10.1</td>
</tr>
<tr>
<td>Control-rel. N</td>
<td>0.20±0.03</td>
<td>31.9±2.1</td>
<td>-6.6±6.2</td>
<td>-55.4±52.0</td>
</tr>
<tr>
<td>Ammonium sulp.</td>
<td>0.65±0.18</td>
<td>92.2±13.4</td>
<td>12.8±1.1</td>
<td>107.4±19.0</td>
</tr>
<tr>
<td>Uran</td>
<td>1.04±0.24</td>
<td>137.9±17.5</td>
<td>-7.8±9.4</td>
<td>-65.5±10.9</td>
</tr>
<tr>
<td>Calcium nitrate</td>
<td>1.54±0.68</td>
<td>202.1±49.7</td>
<td>-29.2±3.2</td>
<td>-244.1±26.8</td>
</tr>
<tr>
<td>Ammonium nitrate</td>
<td>3.70±0.68</td>
<td>476.3±49.6</td>
<td>-21.8±3.6</td>
<td>-181.9±30.5</td>
</tr>
</tbody>
</table>

(1) Lal (2004). N₂O to equivalent CO₂-C = kg ha⁻¹ N₂O-N x 296. CH₄ to equivalent CO₂-C = g ha⁻¹ CH₄-C x 23. Total emission = [Σ (N₂O equiv. CO₂-C + CH₄ equiv. CO₂-C+ Costs CO₂-C) for fertilized soils] – [Σ (N₂O equiv. CO₂-C + CH₄ equiv. CO₂-C) without fertilizer].

Figure 2. Ammonium contents (NH₄⁺-N; a, b), nitrate contents (NO₃⁻-N; c, d) and soil pH values (e, f) for the 15 day period after application of the N fertilizers to a Gleysol (0 to 0.10 m) in the South of Brazil. Vertical bars mean standard deviation of the mean (n=2).
the larger emissions in ammonium nitrate-amended soils than calcium nitrate, in addition to the elevated NO$_3^-$-N contents and WFPS values, the counter ammonium ion possibly favoured the soils biological activity, especially in the presence of great quantity of crop residues with a high C/N ratio, which in turn possibly increased the oxygen consumption and enhanced the soil anaerobiosis condition (Baggs et al., 2003), favouring the N$_2$O generation by denitrification.

The N$_2$O emission on the third day after N application, apart from the treatments with ammonium nitrate and calcium nitrate, had a close relation to the NO$_3^-$-N contents in the soil (Equation 1, p < 0.08), but the inclusion of NH$_4^+$-N contents to a multiple linear regression markedly decreased the significance (Equation 2, p < 0.17). Such results indicate that denitrification was the major process for N$_2$O generation in the current study, where the NO$_3^-$-N content by itself explained 56 % of the N$_2$O emission and the NH$_4^+$-N content in addition to NO$_3^-$-N improved the relationship in only 13 % (R$^2$ = 0.69). Other evidence for the importance of the denitrification process to the N$_2$O emission in this study was the values of coefficients in the multiple linear equation (Equation 2), indicating that the increase in one unit of soil NO$_3^-$-N content promoted increases in N$_2$O emission threefold larger than the increase of one unit of NH$_4^+$-N content (50.9/16.8 = 3.02), considering the other variables steady.

\[
\text{N}_2\text{O}_{3\text{rd day}} (\mu\text{g m}^{-2} \text{ h}^{-1} \text{ N}) = 404.3 + 28.2 \text{NO}_3^-\text{-N} \quad (1)
\]

\[
\text{R}^2 = 0.56; \quad p < 0.08
\]

\[
\text{N}_2\text{O}_{3\text{rd day}} (\mu\text{g m}^{-2} \text{ h}^{-1} \text{ N}) = -930.1 + 16.8 \text{NH}_4^+\text{-N} + 50.9 \text{NO}_3^-\text{-N} \quad (2)
\]

\[
\text{R}^2 = 0.69; \quad p < 0.17
\]

Two main reasons, which do not exclude each other, might explain the absence of relationship between N$_2$O emission and fertilization with ammonium nitrate and calcium nitrate. Firstly, the soil NO$_3^-$-N contents may have surpassed the soil capacity of response in increases of N$_2$O emission under the conditions of that soil. This hypothesis was already conjectured by Moreira & Siqueira (2006), when the authors theorized that intense N$_2$O fluxes can occur independently of nitrate contents in soils with NO$_3^-$-N > 20 mg L$^{-1}$. The second reason is that, for fertilized soils, the N$_2$O generation from denitrification may be hampered by the availability of labile carbon in the soil, irrespective of the nitrate content (Dobbie & Smith, 2003). This aspect is particularly important to calcium nitrate, wherein denitrification is the only process for N$_2$O generation.

Several studies suggest that denitrification is the major process related to N$_2$O emission from soils, mainly under large water content conditions (Dobbie & Smith, 2003; Liu et al., 2007). Under these favourable conditions and in fertilized soils, Batemam & Baggs (2005) estimated that the N$_2$O amount generated from denitrification was fivefold greater than from nitrification. In this context, many studies reported that the use of ammonium- and/or amidic-based N fertilizers instead of nitric-based sources in humid soils or in applications followed by rainfall or irrigation is an effective strategy to decrease the N$_2$O emissions from such soils (Clayton et al., 1997; Maggiotto et al., 2000; Dobbie & Smith, 2003; Jones et al., 2005), in addition to the split rate application of the N fertilizer (Wolschick et al., 2003; COMISSÃO..., 2004).

The relatively small peak intensities of N$_2$O emission from the soils amended with urease inhibitor urea and controlled-release N evidence their potencial as alternative sources efficient to diminish the N$_2$O emissions credited to N fertilizations (Figure 1b), corroborating the previous results of Zaman et al. (2008). The urease inhibitor presence and the restriction from the protective layer against water entrance around the fertilizer postpone and decrease the NH$_4^+$ concentration in the soil, restraining the nitrification and, consequently, the denitrification process. However, oppositely to the expected results, the NH$_4^+$ contents in the soil amended with these N sources were as large as those from the soil amended with normal urea, suggesting that the product did not hamper the urea hydrolysis to NH$_4^+$ at the soil humidity conditions of the experiment. Therefore, the smaller N$_2$O emissions from soils of these treatments were not necessarily due to the effects of the urease inhibitor or due to the controlled-release of N, but were probably attributed to the lesser nitrification rate of NH$_4^+$ in conditions of large soil humidity. Batemam & Baggs (2005) reported a liquid nitrification of NH$_4^+$ of about 1 mg kg$^{-1}$ d$^{-1}$ N in soil with 70 % WFPS. Similarly, Dobbie & Smith (2003) did not observe effects of urease inhibitor on NH$_4^+$ contents in the soil, reporting inclusively greater contents than in normal urea-amended soil for most evaluation events, which resulted in similar N$_2$O emissions. In the case of the controlled-release N, Dobbie & Smith (2003) found smaller, but consistent, NH$_4^+$ concentrations in the soil than the normal urea, which implied a greater emission due to the long term emission. Maggiotto et al. (2000), in a study of controlled-release N for three years, observed that the effect was distinct according to the year. Therefore, in spite of the promised results from the current study regarding the use of urea with urease inhibitor and the controlled-release N, the potential of such products in decreasing N$_2$O emissions must be systematically investigated before their recommendation for large scale.

Concerning the accumulated N$_2$O emission for the period (15 days), the use of nitric-based N fertilizers resulted in the emission of 3.70, 1.54 and 1.04 kg ha$^{-1}$ N for ammonium nitrate, calcium nitrate and Uran, respectively (Table 4), which were larger than the emissions imparted from the other N sources (0.20 to
0.68 kg ha\(^{-1}\) N). The \(\text{N}_2\text{O}\) emissions from nitric-N fertilizers corresponded to 2.46, 1.03 and 0.69 % of the total N applied to the soil for ammonium nitrate, calcium nitrate and Uran, respectively. The application of ammonium sulphate and urea, however, resulted in the loss of only 0.45 and 0.32 %, respectively. These results point that, for conditions favourable to denitrification (large WFPS), the \(\text{N}_2\text{O}\) emissions imparted from the addition of ammonium- or amidic-based fertilizer exclusively were about 50 % smaller than from nitric-based ones. The accumulated \(\text{N}_2\text{O}\) emissions following the application of urease inhibitor and controlled-release N forms were smaller than for ammonium sulphate and urea (Tables 3 and 4). Application of urea+urease inhibitors resulted in accumulated emission of 0.22 % of the total N applied (0.38 kg ha\(^{-1}\) N), while the controlled-release N achieved an accumulated emission of 0.13 % (0.25 kg ha\(^{-1}\) N).

The range of \(\text{N}_2\text{O}\) emission from soils in literature is wide and is strongly affected by the source and rate of fertilization, as well as by climate and soil characteristics (Clayton et al., 1997). The emissions found in the present study, however, can be considered as large, since the relative losses of N through \(\text{N}_2\text{O}\) emission for only 15 days of evaluation are similar to annual losses from soils following application of nitric sources of N fertilizers (0.4 to 3.1 %), ammonium-based fertilizers (0.2 to 0.4 %), amidic-based fertilizers (0.24 to 0.8 %) and with microbial activity inhibitor (0.1 to 0.52 %) (Bouwman, 1996; Maggiotto et al., 2000; Dobbie & Smith, 2003; Jones et al., 2005). One factor that can explain the large \(\text{N}_2\text{O}\) emissions of the current study is the probable anoxic condition of the soil due to its poor drainage exacerbated by the rainfall occurrence in the period subsequent to the N application (Figure 1).

**Soil CH\(_4\) fluxes**

The soil CH\(_4\) fluxes were affected by the sources of N fertilizer applied to the corn, but the magnitude of such fluxes were up to 300 times smaller than those for \(\text{N}_2\text{O}\) (Figures 1b and 3). The CH\(_4\) fluxes ranged from an influx of -30.1 \(\mu\)g m\(^{-2}\) h\(^{-1}\) C to efflux of +32.5 \(\mu\)g m\(^{-2}\) h\(^{-1}\) C (Figure 3). Such values represent the difference between the methanogenesis and methanotrophy processes in the soil. The range in the CH\(_4\) flux rates was thinner than that cited by Veldkamp et al. (1998) in Costa Rica soils under fertilization (-62.5 to +250 \(\mu\)g m\(^{-2}\) h\(^{-1}\) C-CH\(_4\)). In Brazil, flux rates of -20.8 to + 54.2 \(\mu\)g m\(^{-2}\) h\(^{-1}\) C-CH\(_4\) for high lands of the Mid-West region cultivated with non-flooded rice (Metay et al., 2007) and of -40 to +62 \(\mu\)g m\(^{-2}\) h\(^{-1}\) C-CH\(_4\) of corn-cultivated soils in the Southern region (Gomes, 2006) were documented. Despite the relatively small rates of influx of CH\(_4\) in agriculture soils in general, the negative fluxes evidence the occurrence of methanotrophy in such soils and highlight that, even for mineral N-fertilized soils, the agriculture soils have the potential to act as sinks of atmospheric CH\(_4\), through the CH\(_4\) oxidation to CO\(_2\).

The biggest oscillation in CH\(_4\) flux rates in soil were verified in the first week after N application (Figure 3). Rates increased markedly (60 to 194 %) in one day after N application in comparison to the unfertilized soil (Figure 3) and were closely and directly related to the NH\(_4^+\) contents in the soil (Equation 3).

\[
\text{CH}_4\text{ 1st day (}\mu\text{g m}^{-2}\text{ h}^{-1}\text{ C}) = -23.3 + 0.39\text{ NH}_4^+ \quad (3)
\]

\[R^2 = 0.65; \quad p < 0.02\]

The soil NH\(_4^+\) contents were significantly large in this first week of evaluation (> 40 mg kg\(^{-1}\) N in soil) when compared to the control, inclusively in soils that received N fertilizers containing solely nitrate (Figure 2a,b), presumably due to their effect in increasing the rates of soil organic matter mineralization. The relationship between CH\(_4\) fluxes and NH\(_4^+\) contents is attributed to the inhibitory effect of NH\(_4^+\) on the methanotrophy process, responsible for the CH\(_4\) oxidation in the soil (Hütsch, 1998). According to Hütsch (1996), this effect is very effective and decreases larger than 30 % in the CH\(_4\) oxidation rates can be detected only three hours after N application to the soil. The inhibitory mechanism of NH\(_4^+\) on CH\(_4\) influx is complex and involves both the enzymatic competition for the methane monooxygenase enzyme (Kravchencko et al., 2002) and the toxic effect of composts generated during nitrification (hydroxylamine and nitrite) on methanotrophic
microrganisms (Hütsch, 1998). Accordingly to Kravechencko et al. (2002), NH₄⁺ concentrations greater than 40 mg kg⁻¹ N are sufficient to trigger the inhibitory effect on the soil CH₄ oxidation. When the soil NH₄⁺ availability is large, the methanotrophic bacteria activity changes to the NH₄⁺ oxidation in detriment of CH₄ oxidation, because the CH₄ monooxygenase enzyme has low specificity for its substrate (Hütsch, 2001). This behavior determines, temporarily, greater liquid emission of CH₄ from the soil, but the effect can persist for days, mainly if associated to the maintenance of large NH₄⁺ contents in the soil.

From the third day on, in addition to the NH₄⁺ contents, the CH₄ emission was likely impacted by the WFPS, when the values reached 75 % on this date (Figura 1). While NH₄⁺ is responsible for the methanotrophy decrease due to the enzymatic competition previously discussed, the larger values of WFPS make the CH₄ diffusion from the soil difficult and are responsible for the generation of anoxic microsites in the soil, which in turn increase the methanogenesis occurrence (Kessavalou et al., 1998).

After the seventh day of N application, the differences in CH₄ influx/efflux rates in the soil among the N sources decreased and, from this moment, liquid influx predominated, with rates ranging from -4.4 to -10.1 μg m⁻² h⁻¹ C, likely related to the decrease in WFPS (> 60 %) and in soil NH₄⁺ contents (Figures 1a and 3a-b). The decreasing effect on soil CH₄ oxidation is usually short term and completely reversible (Dunfield & Knowles, 1995). In laboratory studies, Hütsch (1998) observed a gradual recomposition in the oxidation capacity after five days of fertilization, joined by the decrease in the soil NH₄⁺ contents. However, in field experiment, Veldkamp et al. (1998) reported the regress of CH₄ oxidation rates to the original values only after three weeks of fertilization. According to Hütsch (2001), if conditions of low temperature, great soil water content and frequent fertilization persist for long term, the decrease in the soil CH₄ oxidation capacity can become irreversible due to damages in the methanotrophic population. This long term effect is mainly detected when oxidation rates of agricultural soils are compared to adjacent soils under natural pastures and forests. In forest soils, CH₄ oxidation rates in general are substantially greater than in agriculture soils due to the absence of N fertilization (Hütsch, 2001).

The mean accumulated CH₄ emission for the N-amended soils was significantly larger (60 %) than for the non-amended soil (Table 3), which summed an influx of -45.1 g ha⁻¹ C in the 15 day period of evaluation (Table 4). The calcium nitrate (-29.2 g ha⁻¹ C) and urea (-26.3 g ha⁻¹ C) application promoted the smallest injurious effects of mineral N fertilizers on the soil CH₄ oxidation and were followed by ammonium nitrate (-21.8 g ha⁻¹ C) and urea with urease inhibitor (-12.4 g ha⁻¹ C). The greatest increases in the CH₄ liquid emission in comparison to the non-amended soil were found for ammonium sulphate (100 %), uran and controlled-release N (75 %) (Table 4).

The increase in the accumulated CH₄ emission for the period had direct and close relation with the soil NH₄⁺ content (Figure 4). Mosier et al. (1991) were the first authors to describe the suppressive effect of NH₄⁺ on methanotrophy, after of the observation that the application of 22 kg ha⁻¹ N as ammonium nitrate decreased the CH₄ influx of a pasture soil in 41 % in comparison to the non fertilized soil. Veldkamp et al. (1998) also reported that ammonium sulphate was the N fertilizer that nourished the biggest decrease in the CH₄ oxidation rate in the soil (60 % in comparison to calcium nitrate and urea).

In addition the great soil NH₄⁺ contents, another factor that can contribute to the increase in CH₄ emission from soils fertilized with ammonium sulphate is the decrease in soil pH. This fertilizer has acid reaction in the soil and, in the present study, promoted a decrease in soil pH in 0.3 units, at average for the period of evaluation (Figure 2e-f). In addition to the direct effect on the methanotrophic population, the pH decrease also prolongs the period rerequired for the nitrification of the NH₄⁺ applied to the soil, as nitrification rates are smaller in low pH soil (Moreira & Siqueira, 2006).

Environmental impact from N fertilizers concerning greenhouse gases emission

The environmental impact of N fertilizers must take into account the N₂O and CH₄ emissions from the soil, as well as the emissions of all GHG (“costs in CO₂”) involved with the manufacture, transport and application of the fertilizers. In table 4, the N₂O and CH₄ emissions from soil are represented in quantities

![Figure 4. Relationship between accumulated flux of CH₄ (g ha⁻¹ C) and mean contents of ammonium (mg N kg⁻¹) in the soil for the 15 day period after application of the nitrogen fertilizers to a Gleysol in the South of Brazil.](image-url)
of equivalent CO$_2$-C based on the global warming potential of N$_2$O (296) and CH$_4$ (23) in comparison to CO$_2$ (IPCC, 2007) and the costs in CO$_2$ of the N fertilizers application (150 kg ha$^{-1}$ N) equivalent to 195 kg ha$^{-1}$ C-CO$_2$, which were estimated from the coefficient of emission of 1.3 kg CO$_2$-C kg$^{-1}$ N, as suggested by Lal (2004).

The magnitude of N$_2$O and CH$_4$ emissions, in equivalent CO$_2$-C, highlights that the environmental impact from N$_2$O emission from soil was markedly larger than that from CH$_4$ emission (Table 4). For the average among the treatments, the liquid CH$_4$ emission was equivalent to 0.2 kg ha$^{-1}$ CO$_2$-C, which corresponded to about 500 times smaller than the mean for N$_2$O emission (132.8 kg ha$^{-1}$ CO$_2$-C). Among the N-amended soils, the N$_2$O emission range was equivalent to the emission of 31.9 to 476.3 kg ha$^{-1}$ CO$_2$-C (Table 4). After summing the emission of both gases and the mean cost of the fertilizers in CO$_2$-C (195 kg ha$^{-1}$ CO$_2$-C), the mineral N fertilization promoted total emissions of 220.4 to 664.5 kg ha$^{-1}$ CO$_2$-C equivalent to atmosphere (Table 4). The nitric-based fertilizers had emissions larger than 326 kg ha$^{-1}$ CO$_2$-C.

Taking into account these values, it is possible to infer that the effect of N fertilization on the enhancement in the soil organic matter stocks due to their increase in biomass production can only partially counterbalance the harmful impact on GHG emissions to the atmosphere (220.4 to 664.5 kg ha$^{-1}$ CO$_2$-C). In studies carried out in the South of Brazil, the uppermost rate of annual C accumulation in soil due to N fertilization was 160 kg ha$^{-1}$ CO$_2$-C (Zanatta et al., 2007; Weber, 2008). Oppositely, however, the results point out that there are possibilities to minimize the N$_2$O emissions decurrent from mineral N fertilization and that, under large water content in soil, this goal can be achieved by using ammonium- and amidic-based fertilizers in detriment of nitric sources. This potential can be even better exploited if controlled-release N and the urea with urease inhibitor are adopted. Complementing these alternatives, it is worthy of mentioning that the adequate moment of application and the adoption of split rates are key strategies for maximizing the efficiency of nutrient use.

CONCLUSIONS

1. The use of N fertilizers intensified the N$_2$O emissions from the Gleysol evaluated in this study. The nitric-based sources promoted larger N$_2$O emissions under soil conditions of large water-filled pore space and denitrification was the main process involved in the gas generation. Ammonium- and amidic-based fertilizers represented better alternatives for decreasing the N$_2$O emission in this poor drained soil, as well as the controlled-release N and the urea with urease inhibitor.

2. Nitrogen fertilization increased the CH$_4$ emission from the soil, but the magnitude of CH$_4$ fluxes in consequence of N fertilization was several hundred times smaller than the N$_2$O fluxes from soil. The environmental impact from such increases in CH$_4$ emissions due to nitrogen fertilization, therefore, was of low significance.

3. The main driving variables controlling the N$_2$O emission in the Gleysol were the nitrate content in soil and the water-filled pore space, while the NH$_4^+$-N content was the main driving variable for the CH$_4$ emission from soil. For both gases, the main injurious effect of N fertilizer application to the soil on their emissions was ephemeral.

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NITROUS OXIDE AND METHANE FLUXES IN SOUTH BRAZILIAN GLEY SOL AS AFFECTED


